

Original Article

Genetic Variability of *Hemiscorpius lepturus* in Khuzestan Province, Iran, Using ISSR-PCR and Mitochondrial Cytochrome C Oxidase Subunit I Gene SequencingKobra Chehari¹, Abbas Jolodar^{1*} , Hediye Jafari² 

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ABSTRACT

Background: Scorpion stings pose a serious public health concern, particularly in Khuzestan Province, Iran. The genus *Hemiscorpius* scorpions are a major cause of scorpion stings and related deaths, mostly in children.

Objectives: This study aimed to study the potential intraspecific variability of *Hemiscorpius lepturus* in Khuzestan.

Methods: We used inter-simple sequence repeat anchored-polymerase chain reaction (ISSR-PCR) and cytochrome c oxidase subunit 1 (COI) to study potential intraspecies variability of this scorpion. Twenty-two specimens of *H. lepturus* scorpions were collected from 5 geographically distinct regions of Khuzestan. Genomic DNA was extracted using the phenol/chloroform method. For phylogenetic analysis, target gene fragments were amplified using ISSR-PCR. By agarose gel electrophoresis of the PCR products, bands produced in each specimen were categorized using a zero and one system, and a dendrogram was drawn using the UPGMA algorithm.

Results: ISSR-PCR generated 5 bands ranging from 0.9 to 2.5 kb. The results showed that the specimen H14Ch was clearly different within its group. To validate these findings, a 637-nucleotide fragment of the COI gene was amplified and sequenced from 5 genetically variable specimens. Out of 5 sequences, H11Ba and H16Be have a relatively close relationship (57%) with the *H. lepturus* reference sequence (KU341987). However, H14Ch was placed with a relatively high distance (72%) from the rest of them, next to the other reference sequence *Hemiscorpius* sp. (OP433762.1).

Conclusion: Although the scorpions of each region were mostly placed together in the phylogeny tree, no major genetic diversity related to regional differences was observed in the province. Based on the genetic distance of H14Ch from other sequences (12.5%), it is definitely an intraspecies variation.

Keywords: Dendrogram, Inter simple sequence repeat (ISSR), Khuzestan, Phylogeny, Scorpion

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Introduction

Over the last two decades, the number of scorpion species, genera, and families worldwide has changed, with nearly 2231 species now described. To date, 2231 scorpion species have been classified into 208 genera and 20 families (Shahbazzadeh et al., 2003). Iran has one of the most diverse scorpion faunas in West Asia. Therefore, it is one of the best areas for studying scorpions (Dehghani et al., 2016). Despite this high species diversity, the scorpion fauna and geography have not been fully studied due to the vast size of the country. According to the latest review of scorpion classification in Iran, there are three families: Buthidae (51 species), Scorpionidae (3 species), and Hemiscorpiidae (6 species), with representatives from different parts of the country reported (Dehghani et al., 2016).

Among the existing scorpions of Khuzestan, the yellow scorpion called Gadim, with a scientific name of *Hemiscorpius lepturus*, is one of the most common and dangerous scorpions in Iran. This scorpion belongs to the family Hemiscorpiidae. Three species of this genus, *Hemiscorpius gailliardi*, *H. lepturus*, and *Hemiscorpius persicus*, have been reported in Iran. The distribution of *H. lepturus* scorpion appears to be limited to Iraq and the western and southwestern regions of Iran. However, the distribution area of this species also includes western Pakistan (Lourenco, 2001). The highest mortality rate from *H. lepturus* stings has been reported in Khuzestan Province, Iran. The venom of this scorpion leads to severe hemolysis and cardiovascular disorders (Radmanesh, 1998).

It is well known that, despite the close morphological similarities, it is difficult to differentiate scorpions using this feature alone (Polis et al., 1990). Therefore, a molecular taxonomic study can be an appropriate solution to this problem. In one study, species boundaries for *Mesobuthus przewalski* from northwestern China were determined using phylogenetic analysis of the mitochondrial marker cytochrome oxidase c subunit 1, ecological modeling, and morphological comparison (Zhang et al., 2019). Other studies on scorpions in Khuzestan Province using mitochondrial genes include scorpion phylogeny of *Ortostirus iranensis* (Jafari et al., 2017), *Mesobuthus eupeus* (Nikkahah et al., 2019), *Odontobuthus* genus (Mirshamsi et al., 2010), and *H. lepturus* (Jolodar, 2019). A study on *Androctonus crassicauda* in Turkey using the cytochrome oxidase gene found no evidence of intraspecific diversity (Oscan et al., 2007).

Phylogenetic studies based on nuclear and mitochondrial genes face limitations due to the need for sequencing, a relatively expensive process. Therefore, in cases where taxonomic studies require a large number of samples and access to genomic DNA is also restricted, using intersimple sequence repeat (ISSR) markers is preferable in mammals and plants due to their simplicity and low cost (Weber et al., 1990). The technique for studying several eukaryotic species was first developed by Zietkiewicz et al. (1994). This method was used to study *Arabidopsis* plants by designing primers targeting repetitive CT nucleotides, using the random amplified microsatellite polymorphism (RAMP) technique (Wu et al., 1994). It has also been used to study fungal genomes (Hantula et al., 1996). The ISSR-polymerase chain reaction (PCR) was used with its concentration on the use of microsatellites that are dispersed in the genome of eukaryotes, to study the intraspecific variability of *Trypanosoma cruzi*, *Leishmania braziliensis*, and *Schistosoma mansoni*, showing that the band profiles obtained were comparable to those resulting from AP-PCR (Oliveira et al., 1997; Uras et al., 2024). A dendrogram using the UPGMA algorithm for scorpions in the *Hottentotta* genus from Khuzestan Province revealed intraspecific genetic diversity (Pirmoradi et al., 2021). Similar results were obtained with SSR-PCR and AP-PCR using isolated *T. cruzi* strains from chronic patients with Chagas disease (Gomes et al., 1998). Given the presence of unknown populations of *H. lepturus* in Khuzestan, we studied potential intraspecific variability in this scorpion.

Materials and Methods

Collection of scorpion specimens

A total of 22 scorpions *H. lepturus* were caught at night using ultraviolet (UV) light from 5 geographically different regions, including the cities of Izeh (3 specimens 49°52'56"N, 31°46'13"E), Baghmalek (7 specimens, 31°54'16"N, 49°20'14"E), Behbahan (3 specimens 30°14'46"N, 50°12'17"E), ChoghaZanbil (5 specimens 32°00'54"N, 48°31'03"E), and Masjedsoleyman (4 specimens 31°46'30"N, 49°00'37"E) (Figure 1). They were identified in the Razi Vaccine and Serum Research Institute in Ahvaz using the identification key (Vachon et al., 1974; Lamoral et al., 1979).

Genomic DNA extraction

To extract genomic DNA, 0.1-0.5 g of scorpion metasoma was crushed in liquid nitrogen, and then 600 μ L of RSB buffer (10 mM Tris-HCl, pH 7.4 / 10 mM NaCl / 25 mM EDTA, SDS 1%) was added, and the mixture

was homogenized. Genomic DNA was extracted with the same amount of phenol/chloroform, then once with the same amount of chloroform. Finally, it was precipitated with pure ethanol and 3 M sodium acetate. DNA concentration and purity were determined by calculating the A260/280 absorbance ratio.

ISSR-PCR amplification

For the ISSR primer 5'-CAA(CT)6, the protocol described by Wu et al. (1994) was followed, except that the annealing temperature was 42 °C. A schematic representation of simple sequence repeat and inter-simple sequence repeat regions is shown in Figure 2. The ISSR-PCR technique relies on primer anchoring at the 3' or 5' ends of microsatellites. To perform the ISSR-PCR reaction, each PCR sample contains 25 µL of 5 ng genomic DNA, 1X PCR buffer, dNTPs (0.25 mM), magnesium chloride (1.5 mM), primer (1 µM), and Taq DNA polymerase (0.5 U). The thermal program for PCR was performed at 95 °C for 3 minutes (one cycle), 94 °C for 45 seconds, 42 °C for 45 seconds, and 72 °C for 60 seconds, with 30 repetitions, and finally 72 °C for 5 minutes. By electrophoresis of the PCR product, gene fragments produced in each specimen were counted and categorized using a zero and one system. DendroUPGMA program (Garcia-Vallve et al., 1999) was used to calculate a similarity index, the Dice coefficient. It performed clustering using the Unweighted Pair Group Method with Arithmetic Mean (UPGMA). The created data was used to draw a dendrogram tree with the help of the program ETAToolkit (Huerta-Cepas et al., 2016).

PCR amplification of cytochrome c oxidase subunit 1 (COI) was carried out on aliquots of the *H. lepturus* genomic DNA as template with initial denaturation for 5 min at 95 °C, followed by 35 cycles of 45 s at 94 °C, 1 min at 48 °C, and 1 min at 72 °C and, finally, 7 min of incubation at 72 °C in a final reaction volume of 25 mL containing 50 mM KCl, 10 mM Tris-HCl (pH 8.3), 0.2 µM of each primer, deoxynucleotides (each at 220 mM), 1.5 mM MgCl₂, 100 ng genomic DNA, and 1 U Taq polymerase. The primers were COI-F 5'-GGTCAACAAATCAT-AAGATATTGG and COI-R 5'- TAAACTTCAGGGT-GACCAAAAAATCA (Folmer et al., 1994). The amplified PCR products were electrophoresed on a 1% agarose gel and stained with DNA Safe Stain (Sinaclon, Iran) before detection under UV transillumination. The amplified fragments were sequenced in both strands using a dideoxy termination method and run on an Applied Biosystems 373 DNA sequencer. The phylogenetic analysis was performed using the neighbor-joining method with 1000 bootstrap replicates in MEGA7.

Results

ISSR-PCR

The quantity and quality of the extracted genomic DNA were estimated to be between 1.56 and 1.73. The DNA concentration of the samples ranged from 8 to 55 ng per microliter. The agarose gel electrophoresis profiles were obtained by ISSR-PCR using 22 *H. lepturus* specimens and the primer CAA(CT)6. The gel electrophoresis profile of a representative set of amplification specimens is shown in Figure 3. In this technique, the DNA sequence is amplified between two microsatellite inverted repeats located at a suitable distance from each other. The bands produced in each lane were compared with those in all other lanes of the same gel. The most easily distinguishable bands were considered for analysis. They were counted and categorized using the binary system (0 and 1). A Dendrogram was constructed based on the presence/absence of each band. Almost all specimens produced bands in the molecular weight range of 0.9-2.5 kb. Amplification with CAA(CT)6 primer in all *H. lepturus* specimens showed a certain pattern, producing a polymorphic band in at least one region. It was the production of a band around 1 kb, which probably indicates a protected band.

A dendrogram of 22 *H. lepturus* scorpion specimens from Khuzestan, based on comparisons of amplified gene fragments, was drawn (Figure 4). The three specimens H12Ba, H110Iz, H13Ch, and H14Ch were distinct, so they showed the greatest separation from the other specimens in their own group. *M. eupeus* formed a separate branch as an out group.

Phylogeny based on COI sequence data

After removing the primer sites, the 637-nucleotide gene fragments named H1Ba, H12Ba, H13Ch, H14Ch, and H16Be were amplified. To compare the amplified nucleotide sequence with sequences in GenBank, a BLASTn search was performed using the highly similar sequences (megablast) option. The target sequence was similar to 12 nucleotide sequences from the Scorpinoidea family, all belonging to the genus *Hemiscorpius*. Among them, three sequences were *H. lepturus* (Table 1).

To compare the target nucleotide sequences, an alignment was performed using the ClustalW program. As shown in Figure 5, the highest number of gaps was observed in the area 33-102. The rest of the areas are almost without gaps. The highest amount of similarity is seen in nucleotide sequences 390-544. The similarity of

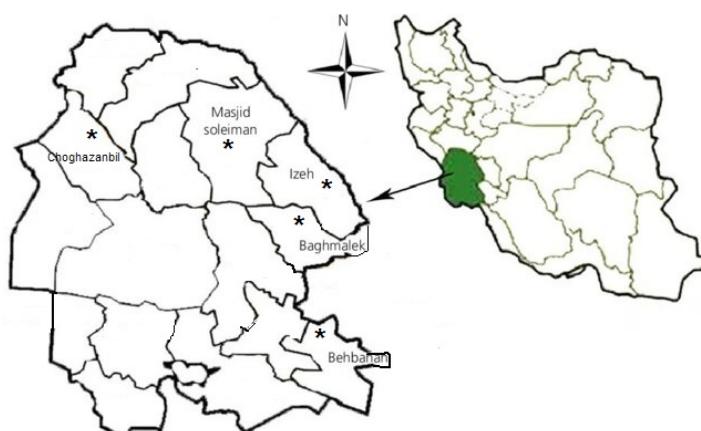


Figure 1. The collecting location of the studied scorpions on a map of Khuzestan Province

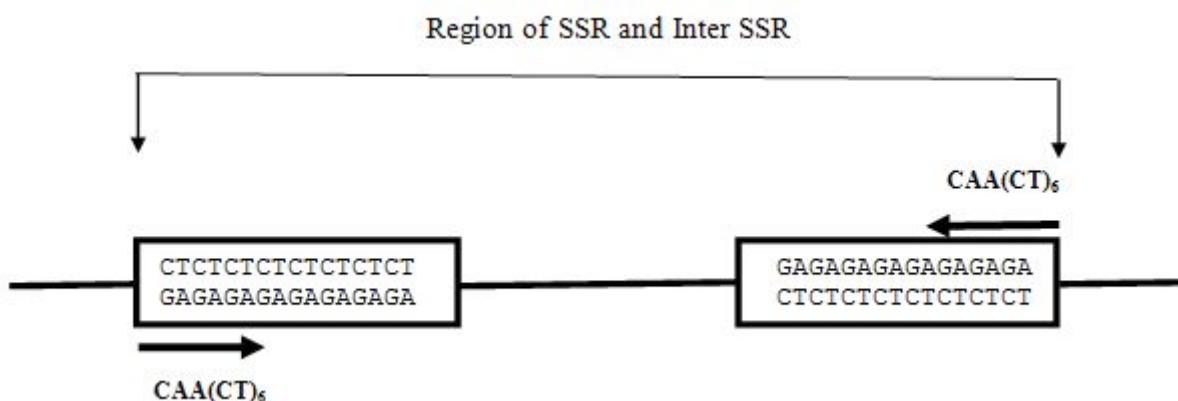


Figure 2. Schematic representation of simple sequence repeat and inter-simple sequence repeats region primer CAA(CT)6

Hl4Ch with the other 4 sequences in the aligned region ranged from 71% to 77%, whereas the similarity among those 4 sequences ranged from 86% to 94%.

Phylogeny based on the *COI* gene

The phylogenetic tree of *H. lepturus* scorpions from Khuzestan was analyzed based on protein and nucleotide sequences of the *COI* gene. The relevant sequences were retrieved from the NCBI database using the BLASTn program. Then, a phylogenetic tree was constructed from *COI* gene sequences of *H. lepturus* scorpions

from Khuzestan using the Neighbor-Joining method in MEGA7. In Figure 6, of the 5 sequences, *Hl1Ba* and *Hl6Be* have a relatively close relationship (57%) with the *H. lepturus* reference sequence (KU341987). *Hl2Ba* and *Hl3Ch* were placed next to each other with a correlation of 89%-90%. *Hl4Ch* was placed with a certain distance (72%) from the rest of them, next to another reference sequence *Hemiscorpius* sp. (OP433762.1). A human sequence (NC012920) is shown as an out-group (Figure 6a).

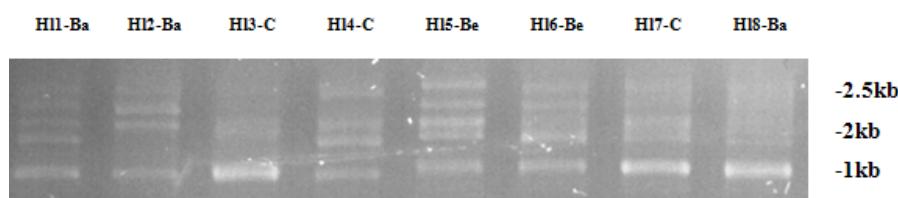


Figure 3. Agarose gel electrophoresis of some specimens as representatives of ISSR-PCR products on 1% gel

Abbreviations: Ba: Bagh malek; Be: Behbahan; Ch: Chogha Zanbil; Ma: Masjed soleiman.

Note: The size of the DNA marker is shown on the right.

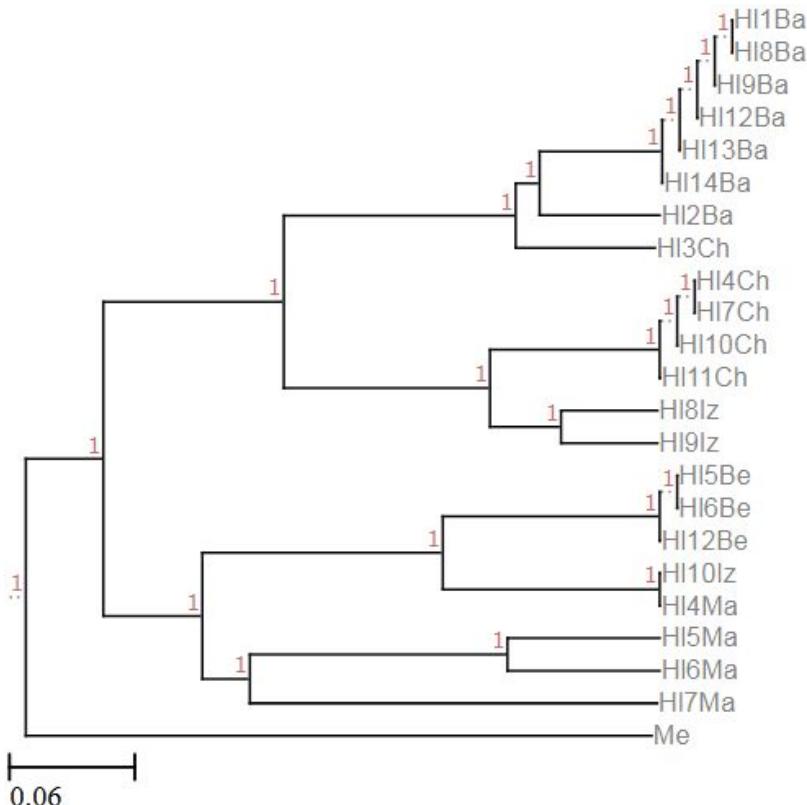


Figure 4. Dendrogram derived from cluster grouping using the band pattern of *H. lepturus* scorpion specimens using the UP-GMA algorithm

Abbreviations: Ba: Baghmalek; Be: Behbahan; Ch: ChoghaZanbil; Ma: Masjedsoleiman.

H11Ba	AGGAGTATTTGG	AACTATATTT	AATTT	AGGA	--GGATGAGC	TCT	--ATGGTGGGACTGCTTAAAG	--ATTAT	--GGTTC	GTC	GTC	--GAAGTA	87					
H12Ba	AGGAGTATTTGG	GACTATGT	TTT	AATTT	--GGATGAGC	TCT	--ATGGTGGGACTGCTTAAAG	--ATTAT	--GGTTC	GTC	GTC	--GAAGTA	88					
H13Ch	AGGAGTATTTGG	GACTATATTT	AATTT	--GGGG	--GGATGAGC	TCT	--ATGGTGGGACTGCTTAAAG	--ATTAT	--GGTTC	GTC	GTC	--GAAGTA	87					
H16Ba	AGGAGTATTTGG	AACTATGT	TTT	AATTT	--GGGG	--GGATGAGC	TCT	--ATGGTGGGACTGCTTAAAG	--ATTAT	--GGTTC	GTC	GTC	--GAAGTA	100				
H14Ch	AGGAGTATTTGG	GGGCTAAGT	TTT	AATTT	GGGG	--GGATGAGC	TCTATA	--ATGGTGGGACTGCTTAAAG	--ATTAT	--GGTTC	GTC	GTC	--GCAC	90				
H11Ba	GGAAACCCGGGG	--CT	TT	ATTTGGGATG	TCAGA	--TTATAATG	TGTGGTT	--CGGCTCA	AGC	TTGTG	ATG	TTT	--GGTATGCCT	ATT	183			
H12Ba	GGAAACCCGGGG	--CT	TT	ATTTGGGATG	ACAGA	--TTATAATG	TGTGGT	--AACGGCTC	AGC	TTGTG	ATG	TTT	--GGTATGCCT	ATT	184			
H13Ch	GGAAACCCGGGG	--CT	TT	ATTTGGGATG	TCAGA	--TTATAATG	TGTGGT	--ACGGCTC	AGC	TTGTG	ATG	TTT	--AGTGTGCTT	ATT	183			
H16Ba	GGAAACCCGGGGGGGCTT	TT	ATTTGGGATG	TCAGA	--TTATAATG	TGTGGT	--ACGGCTC	AGC	TTGTG	ATG	TTT	--AGTGTGCTT	ATT	200				
H14Ch	GTGTTC	CCGGGG	--CC	TT	ATTTGGGATG	TCAGA	--TTATAATG	TGTGGT	--CGGCTC	AGC	TTGTG	ATG	--GGTATGCCT	TTT	186			
H11Ba	ATGATTTGGGG	TT	GGAAATG	TTAGTACCT	TTAAT	TTATGGGAGC	CCTC	--TATGGCTT	TCT	CGT	TTAAATAAC	ATG	TTTGTATT	GGCC	283			
H12Ba	ATGATTTGGGG	TT	GGAAATG	TTAGTACCT	TTAAT	TTATGGGAGC	CCTC	--TATGGCTT	TCT	CGT	TTAAATAAT	ATG	TTTGTATT	GGCC	284			
H13Ch	ATGATTTGGGG	TT	GGAAATG	TTAGTACCT	TTAAT	TTATGGGAGC	CCTC	--TATGGCTT	TCT	CGT	TTAAATAAT	ATG	TTTGTATT	GGCC	283			
H16Ba	ATAATGGGGGG	TT	GGAAATG	TTAGTACCT	TTAAT	TTATGGGAGC	CCTC	--TATGGCTT	TCT	CGT	TTAAATAAT	ATG	TTTGTATT	GGCC	300			
H14Ch	ATGA	--AGGAG	TCAGGCG	TCAGA	TTAGTACCT	CAAA	TTATGGGAGC	CCTC	--TATGGCTT	CCC	CGT	TTAAATAAT	ATRAC	TTTGTATT	GGCC	283		
H11Ba	CGGGCTTTTTCT	TT	TTGGGT	TG	TCG	GC	TTTGGGAAAGAGGGAGC	GGG	GT	GGTTGAAACT	G	GTA	CCAC	CT	TATATCTC	TATATGTT	CACTTCAGG	383
H12Ba	CGGGCTTTTTCT	TT	TTGGGT	TG	CGC	GC	TTTGGGAAAGAGGGAGC	GGG	GT	GGTTGAAACT	G	GTA	CCCG	CT	TATATCTC	TATATGTT	CACTTCAGG	384
H13Ch	CAGGGCTTTTTCT	TT	TTGGGT	TG	CGC	GC	TTTGGGAAAGAGGGAGC	GGG	GT	GGTTGAAACT	G	GTA	CCCG	CT	TATATCTC	TATATGTT	CACTTCAGG	383
H16Ba	CGGGCTTTTTCT	TT	TTGGGT	TG	CGC	GC	TTTGGGAAAGAGGGAGC	GGG	GT	GGTTGAAACT	G	GTA	CCAC	CT	TATATCTC	TATATGTT	CACTTCAGG	400
H14Ch	CGGGCTTTTTCT	TT	TTGGGT	TG	CGC	GC	TTTGGGAAAGAGGGAGC	GGG	GT	GGTTGAAACT	G	GTA	CCAC	CT	TATATCTC	TATATGTT	CACTTCAGG	383
H11Ba	AGGTCTA	GTGGGATAT	GACTATTTT	CTTACATT	GGCTGGGG	TTCTCT	TTATTTGGGGCT	TATTAA	TTT	TTAATAC	T	CTATTCTTA	ATATG	CGCGAGA	483			
H12Ba	TGGTCT	GTGGGATAT	GACTATTTT	CTTACATT	GGCTGGGG	TTCTCT	TTATTTGGGGCT	TATTAA	TTT	TTAATAC	T	CTATTCTTA	ATATG	CGCGAGA	484			
H13Ch	TGGTCT	GTGGGATAT	GACTATTTT	CTTACATT	GGCTGGGG	TTCTCT	TTATTTGGGGCT	TATTAA	TTT	TTAATAC	T	CTATTCTTA	ATATG	CGCGAGA	483			
H16Ba	TGGTCT	GTGGGATAT	GACTATTTT	CTTACATT	GGCTGGGG	TTCTCT	TTATTTGGGGCT	TATTAA	TTT	TTAATAC	T	CTATTCTTA	ATATG	CGCGAGA	500			
H14Ch	TGGTCT	GTGGGATAT	GACTATTTT	CTTACATT	GGCTGGGG	TTCTCT	TTATTTGGGGCT	TATTAA	TTT	TTAATAC	T	CTATTCTTA	ATATG	CGCGAGA	483			
H11Ba	GATGG	ATAGTTTGGATC	GTC	CCCT	TGTTGTT	GATCGGTT	AAAGGT	TACT	GCA	G	G				544			
H12Ba	GATGG	ATAGTTTGGATC	GTC	CCCT	TGTTGTT	GATCGGTT	AAAGGT	TACT	GCG	G	G				545			
H13Ch	GATGG	ATAGTTTGGATC	GTC	CCCT	TGTTGTT	GATCGGTT	AAAGGT	TACT	GCA	G	G				544			
H16Ba	GATGG	ATAGTTTGGATC	GTC	CCCT	TGTTGTT	GATCGGTT	AAAGGT	TACT	CCG	G	G				561			
H14Ch	GATGG	ATAGTTTGGATC	GTC	CCCT	TGTTGTT	GATCGGTT	AAAGGT	TACT	CCG	G	G				544			

Figure 5. Nucleotide sequence comparison of 5 *H. lepturus* scorpion from Khuzestan Province based on the mitochondrial nucleotide sequence of the *COI* gene

Note: Identical nucleotides are shown in dark color. Conserved nucleotides are shown in gray.

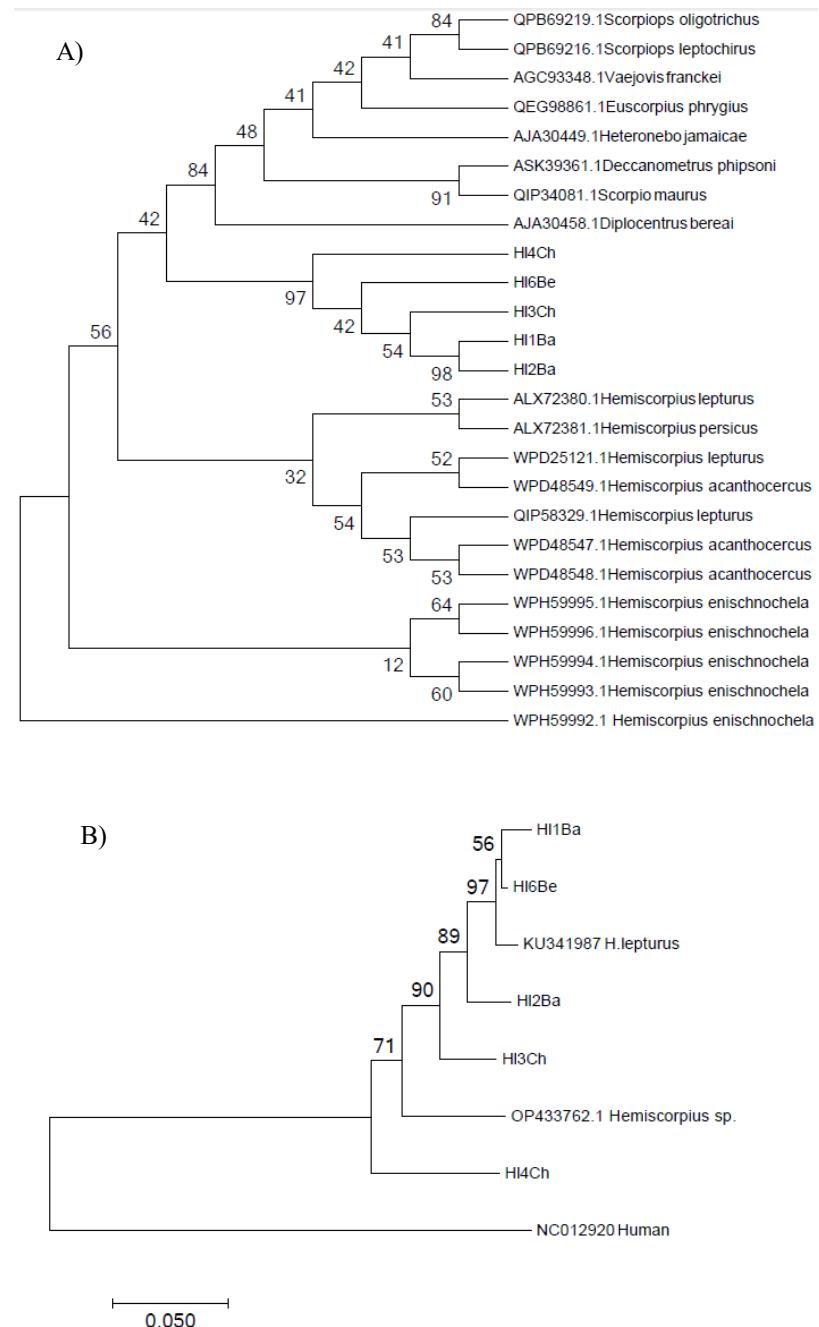


Figure 6. Phylogenetic tree of five *H. lepturus* scorpions from Khuzestan based on A) Nucleotide sequence and B) Protein sequence of the *COI* gene using neighbor-joining analysis

Note: Bootstrap numbers are based on 1000 replicates. The numbers before the species are the accession numbers of the related genes in GenBank. The numbers above the lines indicate the relationship between the groups.

A phylogeny tree of five *H. lepturus* scorpions from Khuzestan, based on the *COI* protein sequence, was constructed and compared with similar sequences from other scorpions (Figure 6b). The results showed that the protein sequences of Khuzestan scorpions were closely related to the scorpion branch within the Euscorpiinae and Diplocentridae families (84%). The 12 reference sequences of *Hemiscorpius* scorpion were grouped into a

sub-branch with a moderate correlation (56%). Three of them are related to *H. lepturus*.

In a brief comparison of the phylogenetic trees obtained from protein and nucleotide sequences, we find that the percentage correlation between the nucleotide sequences of *H. lepturus* specimens from Khuzestan and the two sequences available in NCBI GenBank

Table 1. Taxonomy report of the *COI* gene fragment based on the BLASTn program

Organism	BLAST Name	Score	Number of Hits
<i>Hemiscorpius</i>	Scorpions		12
<i>H. lepturus</i>	Scorpions	161	3
<i>Hemiscorpius acanthocercus</i>	Scorpions	151	3
<i>Hemiscorpius enischnochela</i>	Scorpions	149	5
<i>H. persicus</i>	Scorpions	148	1

(KU341987) ranges from 57% to 72%. Still, this degree of similarity with the 12 protein sequences of the genus *Hemiscorpius* available in Genbank is 56%. Among the 12 reference sequences, three belong to *H. lepturus* and were isolated from Iran (ALX72380 and WPD25121) and Iraq (QIP58329).

Genetic distance

The genetic distance of five *H. lepturus* scorpion specimens from Khuzestan was calculated using the MEGA7 program, compared with each other and with only two *H. lepturus* sequences available in the gene bank (Table 2). The genetic distance of all Khuzestan scorpion specimens, except H4Ch, ranged from 1.6% to 6.6%. However, the H4Ch showed the greatest distance from other sequences (12.5%). The lowest genetic distance (1.6%) to the reference sequence (KU341987) was observed for Hl6B.

Discussion

Scorpions of the genus *Hemiscorpius* are considered to be an important cause of scorpion stings and the result-

ing deaths, especially in children. The Hemiscorpionidae family, with more than 4 species, is among the deadliest scorpion families in Khuzestan Province (Shahbazzadeh et al., 2003). Today, species identification has shifted from morphology to genetic and evolutionary aspects.

Using the ISSR-PCR technique, it is possible to analyze genomic DNA sequences for the taxonomy of various organisms, including scorpions, without prior information. Therefore, the use of primers containing repetitive sequences, which are generally found in intergenic regions and introns in arthropods, has been recommended (Toth et al., 2000). Such primers are not only able to distinguish closely related species but also to distinguish between different populations (De Leon et al., 2010). In this study, molecular phylogeny was evaluated using microsatellite markers in the scorpion *H. lepturus* in Khuzestan Province.

Although the taxonomy of all 22 studied scorpions was identified as *H. lepturus* based on morphological data, genetic differences were observed in both Hl3Ch and Hl4Ch, with this variation more pronounced in Hl4Ch. This level of intraspecies diversity was unexpected given

Table 2. Genetic distance of *H. lepturus* scorpion from Khuzestan using *COI* nucleotide sequence

Organisms	1	2	3	4	5	6	7	8
Hl1Ba								
Hl2Ba	0.051							
Hl3Ch	0.066	0.055						
Hl4Ch	0.125	0.113	0.102					
Hl6Be	0.016	0.035	0.051	0.117				
KU341987_ <i>H. lepturus</i>	0.023	0.039	0.059	0.121	0.016			
OP433762.1_ <i>Hemiscorpius sp.</i>	0.094	0.090	0.090	0.125	0.090	0.102		
NC012920_ Human	0.414	0.422	0.406	0.406	0.414	0.414	0.398	

the presence of a sexual cycle in scorpion reproduction. Therefore, it is possible that this level of genetic diversity, as suggested in the case of the snail genus *Biomphalaria* (Paraense et al., 1957; Jarne et al., 1991), is due to other effective factors, such as recombination, mutation, or gene flow. However, this difference may also be an interspecies variation. To investigate the possibility observed among the studied scorpions, we decided to use the *COI* gene to confirm genetic differences and their geographical distribution in the province.

The genetic distance of all specimens from Khuzestan ranged from 1.6% to 6.6%, except for H14Ch. The results show that specimen H14Ch was dissimilar within its group. This specimen showed the highest distance (12.5%) among others. According to the threshold limit for interspecies genomic variation, suggested to be 13.4% in invertebrates (Kumar et al., 2017), the difference is insufficient to warrant classification as a new species, but it is clearly intraspecific variation.

Geographical and climatic features indicate that Khuzestan Province is divided into two regions: *mountainous* and plain. The low-altitude plain regions of Khuzestan include Behbahan, which accounts for approximately 60% of the entire province, while the mountainous region, mainly in the north and east of the province, including Baghmalek, covers about two-fifths of the entire province. The specimens were collected from these two regions. Considering the obvious environmental differences in Khuzestan Province, such as temperature, humidity, and altitude, it was expected that the geographic location of the specimens would be related to their genetic distance, but this was not the case. In fact, the genetic distance between the two Baghmalek (mountainous region) specimens H11Ba and H12Ba was 5.1%. Still, the genetic distance of those two Baghmalek specimens and H16Be from Behbahan (plain region) was 1.6% and 3.5%, respectively.

Conclusion

Although the scorpions of each region were mostly grouped in the phylogeny, no major genetic diversity associated with regional differences was observed within the province. Based on the genetic distance results, it can be concluded that H14Ch is definitely an intraspecies variation. Phylogeny using other genes and more specimens can achieve more accurate results.

Ethical Considerations

Compliance with ethical guidelines

There were no ethical considerations to be considered in this research.

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Authors' contributions

All authors equally contributed to preparing this article

Conflict of interest

The authors declared no conflict of interest.

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References

- Dehghani, R., Motevali Haghi, F., Mogaddam, M., Sedaghat, M. M., & Hajat, H. (2016). Review study of scorpion Classification in Iran. *Journal of Entomology and Zoology Studies*, 4(5), 440-444. [\[Link\]](#)
- De León, J. H., Neumann, G., Follett, P. A., & Hollingsworth, R. G. (2010). Molecular markers discriminate closely related species *Encarsia diaspadicola* and *Encarsia berlesei* (Hymenoptera: Aphelinidae): Biocontrol candidate agents for white peach scale in Hawaii. *Journal of Economic Entomology*, 103(3), 908–916. [\[DOI:10.1603/EC09316\]](#) [\[PMID\]](#)
- Folmer, O., Black, M., Hoeh, W., Lutz, R., & Vrijenhoek, R. (1994). DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. *Molecular Marine Biology and Biotechnology*, 3(5), 294–299. [\[PMID\]](#)
- Garcia-Vallve, S., Palau, J., & Romeu A. (1999). Horizontal gene transfer in glycosyl hydrolases inferred from codon usage in *Escherichia coli* and *Bacillus subtilis*. *Molecular Biology and Evolution*, 16(9), 1125-1134. [\[DOI:10.1093/oxfordjournals.molbev.a026203\]](#) [\[PMID\]](#)
- Gomes, M. L., Macedo, A. M., Pena S. D., & Chiari, E. (1998). Genetic relationships between *Trypanosoma cruzi* strains isolated from chronic chagasic patients in southern Brazil as revealed RAPD and SSR-PCR analysis. *Acta Tropica*, 69(2), 99–109. [\[DOI:10.1016/s0001-706x\(97\)00122-8\]](#) [\[PMID\]](#)

Hantula, J., Dusabenyagasaniy, M., & Hamelin, R. C. (1996). Random amplified microsatellites (RAMS) - a novel method for characterizing genetic variation within fungi. *European Journal of Forest Pathology*, 26(3), 159-166. [DOI:10.1111/j.1439-0329.1996.tb00720.x]

Huerta-Cepas, J., Serra, F., & Bork, P. (2016). ETE 3: Reconstruction, Analysis, and Visualization of Phylogenomic Data. *Molecular biology and evolution*, 33(6), 1635-1638. [DOI:10.1093/molbev/msw046] [PMID]

Jafari, H., Saalabi, F., Jelodar, A., Navidpour, S., Jahanifard, E., Forouzan A. & Masihipour B. (2018). Phylogenetic study on Orthochirus iranus by using morphological and molecular methods (Scorpiones: Buthidae). *Journal of Entomology and Zoology Studies*, 6(3), 304-309. [Link]

Jarne, P., & Delay, B. (1991). Population genetics of freshwater snails. *Trends in Ecology & Evolution*, 6(12), 383-386. [DOI:10.1016/0169-5347(91)90158-T] [PMID]

Jelodar, A. (2019). Molecular Characterization and Phylogeny Analysis Based on Sequences of Cytochrome Oxidase gene From Hemiscorpius lepturus of Iran. *Iranian Journal of Veterinary Medicine*, 13(1), 59-67. [Link]

Jelodar, A., Ezzati, G. M., Jafari, H., & Farzi, M. (2022). Phylogenetic Relationships of Scorpion Compsobuthus matthiesseni Based on Sequences of Internal Transcribed Spacer 2 Gene from Khuzestan Province, Iran. *Archives of Razi Institute*, 77(1), 65-72. [DOI:10.22092/ari.2021.355059.1662] [PMID]

Kumar, V., Sharma, N., & Sharma, A. (2017). DNA barcoding of the Indian blackbuck (Antilope cervicapra) and their correlation with other closely related species. *Egyptian Journal of Forensic Sciences*, 7, 31. [DOI:10.1186/s41935-017-0034-6]

Lamoral, B. H. (1979). The scorpions of Namibia. *Annals of the Natal Museum*, 23, 497-784. [Link]

Lourenco, W. R. (2001). The scorpion families and their geographical distribution. *Journal of Venomous Animals and Toxins*, 7(1), 03-23. [DOI:10.1590/S0104-79302001000100002]

Mirshamsi, O., Sari, A., Elahi, E., & Hosseinie, S. (2010). Phylogenetic relationships of Mesobuthus eupeus (C.L. Koch, 1839) inferred from COI sequences (Scorpiones: Buthidae). *Journal of Natural History*, 44(47-48), 2851-2872. [DOI:10.1080/0022293.2010.512400]

Nikkhah, N., Jelodar, A. & Taghavi Moghadam A. (2018). Phylogenetic analysis of cytochrome oxidase subunit 1 from the Mesobuthus eupeus (Scorpions: Buthidae) of Khuzestan province. *Iranian Veterinary Journal*, 14(3), 102-111. [DOI:10.22055/ivj.2018.63587.1814]

Oliveira, R. P., Macedo, A. M., Chiari, E., & Pena, S. D. (1997). An alternative approach to evaluating the intraspecific genetic variability of parasites. *Parasitology Today (Personal ed.)*, 13(5), 196-200. [DOI:10.1016/S0169-4758(97)01044-2] [PMID]

Oscan, O., Adiguzel, S., Kar, S., Kurt, M., Yakistiran, S., & Cesaretti, Y., et al. (2007). Effects of Androctonus crassicauda (Olivier, 1807) (Scorpiones: Buthidae) venom on rats: Correlation among acetyl cholinesterase activities and electrolytes levels. *Journal of Venomous Animals and Toxins including Tropical Diseases*, 13(1), 69-81. [DOI:10.1590/S1678-91992007000100005]

Paraense, W. L., & Deslandes, N. (1957). Biomphalaria boissyi, synonyme probable de Taphius nigricans. *Annales de Parasitologie Humaine et Comparee*, 32(5/6), 482-90. [Link]

Pirmoradi, S., Jelodar, A., & Jafari, H. (2021). Genetic Diversity of Hottentotta Zagrosensis and H. saulcyi (Scorpions: Buthidae) using RAMS (Random Amplified Microsatellites) in Khuzestan. *Iranian Veterinary Journal*, 17(2), 38-50. [Link]

Polis, G. A. (1990). *The Biology of Scorpion*. Stanford: Stanford University Press. [Link]

Radmanesh, M. (1998). Cutaneous manifestations of the Hemiscorpius lepturus sting: A clinical study. *International Journal of Dermatology*, 37 (7), 500-507. [DOI:10.1046/j.1365-4362.1998.00386.x] [PMID]

Shahbazzadeh, D., Amirkhani, A., Djadid, N. D., Bigdeli, S., Akbari, A., & Ahari, H., et al. (2009). Epidemiological and clinical survey of scorpionism in Khuzestan province, Iran (2003). *Toxicon: Official Journal of the International Society on Toxicology*, 53(4), 454-459. [DOI:10.1016/j.toxicon.2009.01.002] [PMID]

Toth, G., Gaspari, Z., & Jurka, J. (2000). Microsatellites in different eukaryotic genomes: Survey and analysis. *Genome Research*, 10(7), 967-81. [DOI:10.1101/gr.10.7.967] [PMID]

Uras, M. E., Filiz, E., Sen, U., & Ozyigit, I. (2024). Genetic diversity and phylogenetic analysis of Robinia pseudoacacia L. populations using ISSR markers, ITS1 and trnL-F intergenic spacer sequences. *Journal of Forest Science*, 70(1), 1-13. [DOI:10.17221/95/2023-jfs]

Vachon, M. (1974). [Study of the characteristics used to classify the families and genera of Scorpions (Arachnids). 1. Trichobothriotaxis in arachnology. Trichobothrial abbreviations and types of trichobothriotaxis in scorpions (French)]. *Bulletin du Museum National d'Histoire Naturelle*, 140(104), 857-958. [DOI:10.5962/p.272660]

Weber, J. L. (1990). Informativeness of human (dC-dA)n (dG-dT)n polymorphisms. *Genomics*, 7(4), 524-530. [DOI:10.1016/0888-7543(90)90195-Z]

Wu, K. S., Jones, R., Danneberger, L., & Scolnik P. A. (1994). Detection of microsatellite polymorphisms without cloning. *Nucleic Acids Research*, 22(15), 3257-3258. [DOI:10.1093/nar/22.15.3257] [PMID]

Zhang, X. S., Liu, G. M., Zhang, D. X., & Shi, C. M. (2019). Genetic analysis and ecological niche modeling delimit species boundary of the Przewalski's scorpion (Scorpiones: Buthidae) in arid Asian inland [Preprint]. [DOI:10.1101/652024]

Zietkiewicz, E., Rafaeski, A., & Labuda, D. (1994). Genome Fingerprinting by simple sequence repeat (SSR)-anchored polymerase chain reaction amplification. *Genomics*, 20(2), 176-183. [DOI:10.1006/geno.1994.1151]

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