Amelioration of Lipid Peroxidation and Antioxidant Enzymes Status in the Serum and Erythrocytes of Phenylhydrazine-Induced Anemic Male Rats: The Protective Role of Artichoke Extract (*Cynara scolymus* L.)

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Abstract

BACKGROUND: Hemolytic anemia is a disorder characterized by the premature erythrocytes destruction. Phenylhydrazine (PHZ) induces oxidative stress and reactive oxygen species (ROS) formation, which causes hemolytic anemia. *Cynara scolymus* due to its antioxidant compounds, has been used for various therapeutic purposes in traditional medicine.

OBJECTIVES: The present study was designed to evaluate the effects of *Cynara scolymus* extract on PHZ -induced anemia in male rats.

METHODS: Hemolytic anemia was induced by intraperitoneal injection of PHZ (40 mg/kg) for 2 days. Thirty male Wistar rats were divided into five groups. Group 1 (normal control). Group 2 (anemic control) received only PHZ. The groups 3 to 5 were injected with 100, 200, 400 mg/kg of the *Cynara scolymus* by gavage, respectively, daily from day 2 to day 15 after PHZ administration. At the end of the treatment period, blood samples were collected to assess hematological parameters, malondialdehyde (MDA) level and antioxidant enzymes activity, including superoxide dismutase (SOD) and total antioxidant capacity (TAC) in the serum and erythrocytes.

RESULTS: In anemic rats, serum and erythrocytes MDA level increased, but SOD and TAC activity decreased significantly when compared with control group ($P \le 0.05$). These changes were ameliorated by treatment with *Cynara scolymus* at different doses ($P \le 0.05$). Also, improvement in several hematological parameters was observed in anemic rats after administration of *Cynara scolymus* ($P \le 0.05$).

CONCLUSIONS: *Cynara scolymus* extract exhibits protective property against PHZ-induced oxidative stress presumably due to antioxidative activity.

KEYWORDS: Cynara scolymus, Hemolytic Anemia, Oxidative Stress, Phenylhydrazine, Rat

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Introduction

Anemia is defined as a condition in which the number of red blood cells (RBCs) or their oxygen-carrying capacity is inadequate to meet the physiological needs of the body (Prasad *et al.*, 2018). There are several kinds of anemia classified by a variety of underlying causes. Hemolytic anemia is the most frequent form of anemia (Lee *et al.*, 2012). Hemolytic anemia is categorized as acquired or hereditary. Common acquired causes of hemolytic anemia are autoimmunity, microangiopathy, infection and drug (Dhaliwai *et al.*, 2004; Paul *et al.*, 2018).

Phenylhydrazine (PHZ) is a strong oxidant agent which has toxicity on various tissues at various levels. It is known to shorten life-span of erythrocytes resulting in hemolytic anemia, increased erythropoietic activity, enhanced iron absorption and tissue iron overload (Luangaram et al., 2007). PHZ has been used experimentally for the induction of hemolytic anemia in animal models. The auto-oxidation of PHZ leads to generation of ROS and PHZ-derived radicals, which causes a wide variety of deleterious cellular response including hemolytic anemia (Sung et al., 2013). The observed haematotoxicity is a result of the reaction of PHZ with oxygenated hemoglobin to form oxygen radicals and methemoglobin (Shetlar and Hill, 1985).

Medicinal plants and natural products have been used as traditional treatments for numerous diseases particularly in developing countries because of several reasons including therapeutic effects, affordability, accessibility and fewer side effects (Asase *et al.*, 2008). Several studies have demonstrated that natural medicinal plants with potent antioxidant activity and their potential protective effects can alleviate the damage of oxidative stress-associated diseases through inhibition of ROS generation and improvement of antioxidant defense mechanisms (Forni *et al.*, 2019; Banerjee *et al.*, 2018).

Cynara scolymus L. (Artichoke), a member of Asteraceae family, is an ancient herbaceous perennial plant, originating from the Mediterranean area, which today is widely cultivated all over the world because of its nutritional benefits and medicinal purposes (Salekzamani et al., 2019). Phytochemicals analysis of Cynara scolymus has been found to contain powerful polyphenolic compounds which have therapeutic options including remarkable antioxidant activity against ROS and preventing the formation of free radicals (El-Boshy et al., 2017). Previous studies have reported that artichoke extract has important activities such as hepatoprotective (Gebhardt and Fausel, 1997), hypoglycemic (Salem et al., 2017), antibacterial (Shimoda et al. 2003), antioxidant (Salekzamani et al., 2019), anti-inflammatory and immunomodulatory (El-Boshy et al., 2017).

There is little evidence to indicate artichoke is useful to alleviate hemolytic anemia. Therefore, this study was undertaken to evaluate the putative antioxidant action of *Cynara scolymus* extract in an experimental model of PHZ-induced toxicity in Wistar rats.

Materials and methods

Chemicals and preparation of extract

The ethanolic extract of artichoke that was used in this study was purchased from Dineh Iran Industries Complex (Pharmaceutical Company, Tehran, Iran). Phenylhydrazine (PHZ) was purchased from Sigma-Aldrich (St. Louis, MO, USA).

Animals housing conditions

This study was carried out on 30 mature male Wistar rats, weighing approximate-

ly 210-220 gr that were obtained from the Animal Care Unit of Faculty of Veterinary Medicine, Razi University, Kermanshah, Iran. The rats were acclimatized for approximately one week before use. The animals were housed in stainless steel cages under controlled environmental conditions with temperature (22±2 °C), relative humidity of 55±5% and lighting (12-h light/12-h dark cycle). All rats were fed with a standard laboratory pelleted chow diet and fresh water ad libitum. Animal experiments used in this study were approved by the Animal Ethics Committee of Razi University following the Guidelines for the Care and Use of Laboratory Animals in Research (Animal Ethical Approval Number: 397-2-008).

Experimental procedure

After one week of acclimatization, the rats were randomly assigned into five groups with six animals per group. In group 1, animals were orally and daily injected with normal saline and served as the normal control. In groups 2 to 5, anemia was induced by Intraperioneal (IP) injection of PHZ at 40 mg/kg for 2 consecutive days. Rats that developed anemia with hemoglobin (Hb) concentration lower than 14 g/dl were used for the study. The anemic rats were randomly divided into four groups and treated as follows: In group 2, rats were given normal saline orally and served as the anemic control. Groups 3, 4 and 5, anemic rats daily received the hydroalcoholic extract of C. scolymus at doses of 100, 200, 400 mg/kg body weight by oral gavage, respectively. The normal saline and hydroalcoholic extract of C. scolymus was administered from days 2 to 15 after PHZ injection. The experimental period was 14 days. In this study, dose of PHZ to induce anemia and C. scolymus doses were determined according to previous studies (Diallo et al., 2008; Lee et al., 2012; Salem et al., 2017).

Sample collection

At the end of the study period, all rats were weighed and then anesthetized using diethyl ether. Blood samples were taken from the heart to determine the hematological and antioxidant parameters. For erythrocyte preparation, the erythrocytes were sedimented by centrifuge at 500 g for 10 min at 4 °C. The erythrocytes were washed three times (5 ml, each) with cold isotonic saline then the buffy coat was discarded. RBCs and sera were stored at -20 °C until assayed.

Determination of hematological parameters

For hematological analyses, blood was examined using an automatic hematology analyzer (Celltac, Alpha Vet MEK-6550; Nihon Kohden Co, Tokyo, Japan).

Measurement of total antioxidant capacity (TAC) levels in serum and erythrocytes

Spectrophotometer analysis with the aid of colorimetric assay kit (NaxiferTM, Navand-salamat Co., Iran) was used to estimate the concentrations of TAC in RBCs and serum by the ferric reducing ability (FRAP) method. This procedure is based on the ability of serum or RBC lysis to reduce iron III (Fe³⁺) to iron II (Fe²⁺) in the presence of 2,4,6-Tripyr-idyl-S-triazine (TPTZ). A complex with blue color and maximum absorbance appeared at 593 nm with reaction of Fe²⁺ and TPTZ. The serum level of TAC was expressed in nanomoles per milliliter (nmol/ml) and nmol per gr of hemoglobin (nmol/grHb) for RBC lysis.

Measurement of malondialdehyde (MDA) levels in serum and erythrocytes

The levels of lipid peroxidation in the serum and RBCs lysis were measured as TBARS using a Nalondi[™] assay kit (Navandsalamat Co., Iran). Sera were assayed directly using the kit. The RBCs were first Lysed in deionized water containing butylated hydroxytoluene (BHT) provided in the kit,

and were subsequently centrifuged at 10,000 g for 5 min to collect the supernatant. Serum MDA level was expressed as nmol/ml and it is also expressed in nmol/gr Hb in RBCs.

Measurement of superoxide dismutase (SOD) levels in serum and erythrocytes

Copper, zincsuperoxide dismutase (SOD) was assessed by the pyrogallol assay of Nasdox[™] kit (Navandsalamat Co., Iran). The method depends on the spontaneous autoxidation of pyrogallol at alkaline pH, resulting in the production of superoxide anion radicals which cause enhanced autoxidation of pyrogallol. Autoxidation is manifested as an increase in absorbance at 420 nm. Specific activity of SOD was calculated as units/ml (unit/gr Hb for RBCs), in which one unit of enzyme gave 50% inhibition of pyrogallol autoxidation.

Statistical analysis

The obtained data were analyzed by the One-way Analysis of Variance (ANOVA) and Tukey's HSD post-hoc test using IBM SPSS software version 21.0 for Windows. The results were expressed as mean values±SD. The level of statistical significance was set at $P \le 0.05$.

Results

Effect of *cynara scolymus* extract on hematological parameters

The effects of Cynara scolymus extract on hematological parameters in anemic rats are presented in Table 1. The PHZ group showed significant decrease in PCV, Hb, RBC, MCV and MCHC levels compared with the control group ($P \le 0.05$). Administration of HECS (100-400 mg/kg) significantly improved the decreased levels of above mentioned parameters (mainly at doses of 200 and 400 mg/kg) compared to PHZ-treated rats ($P \le$ 0.05). There was also a significant increase in levels of WBC and lymphocyte in PHZ group when compared with control group ($P \le 0.05$). Treatment with artichoke extract only at 400 mg/kg has an increasing impact on WBC level compared to PHZ-exposure group ($P \le 0.05$). Furthermore, we did not observe significant differences in neutrophil levels among all groups (P > 0.05).

Hematological Parameters	Control	PHZ	PHZ+HECS (100 mg/kg)	PHZ+HECS (200 mg/kg)	PHZ+HECS (400 mg/kg)
PCV (%)	41.5 ± 2.11	$28.2 \pm 3.27*$	29.7 ± 1.28	32.3 ± 2.29	$38.8\pm2.96^{\#}$
Hb (gr/dl)	14.2 ± 1.62	$9.4 \pm 1.36*$	10.2 ± 1.71	11.8 ± 1.26	$12.6\pm2.16^{\#}$
RBC (×10 ⁶ /µL)	6.3 ± 0.87	$3.9\pm0.52\texttt{*}$	4.2 ± 0.88	4.8 ± 1.15	$5.9\pm0.73^{\#}$
MCV (fl)	87.4±2.71	60.2±3.10*	68.3±2.01#	80.2±2.34 [#]	85.2±4.55#
MCHC (gr/dl)	32.1±0.75	26.6±1.15*	27.6±0.60	29.8±0.85 [#]	29.5±1.35#
WBC (×10 ³ /µL)	10.2±1.70	13.4±1.35*	13.5±1.05	13.7±1.25	15.0±1.00 [#]
Neutrophil (×10 ³ /µL)	1.4±0.35	1.2±0.25	1.5±0.20	2.2±0.50	2.5±0.85
Lymphocyte (×10 ³ /µL)	7.9±0.50	11.4±1.95*	12.2±1.05	10.6±1.00	11.7±1.05

Table 1. Effect of Cynara scolymus Extract on Hematological Parameters in PHZ-Induced Anemic Rats

Data are expressed as mean \pm SD (n=6). * $P \leq 0.05$ as compared with control group within each row; # $P \leq 0.05$ as compared with PHZ \neg -treated group within each row. PHZ: Phenylhydrazine; HECS: Hydroethanolic Extract of *Cynara scolymus*; PCV: Packed Cell Volume; Hb: Hemoglobin; RBC: Red Blood Cell; MCV: Mean Corpuscular Volume; MCHC: Mean Corpuscular Hemoglobin Concentration; WBC: White Blood Cell.

Effect of *cynara scolymus* extract on TAC levels in serum and erythrocytes

PHZ-induced anemic rats showed a significant reduction of serum and erythrocytes TAC level as compared to the healthy control group ($P \le 0.05$). Administration of *Cynara scolymus* extract at doses of 200 mg/kg and 400 mg/kg considerably increased these parameters toward the control value ($P \le 0.05$). Significant difference in the TAC level between the groups receiving the HECS in anemic rats induced with PHZ was also observed in the current study ($P \le$ 0.05) (Figure 1 and Figure 2).

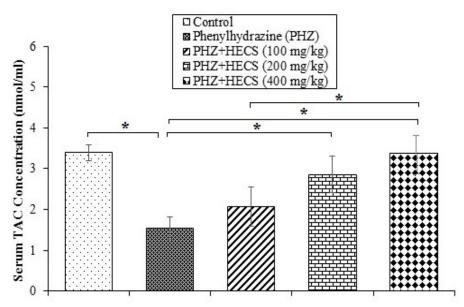


Figure 1. Effect of *Cynara scolymus* extract at different doses on serum TAC concentration in PHZ-induced anemic rats. Data are expressed as mean \pm SD (n=6). * *P* \leq 0.05. PHZ: Phenylhydrazine; HECS: Hydroethanolic Extract of *Cynara scolymus*

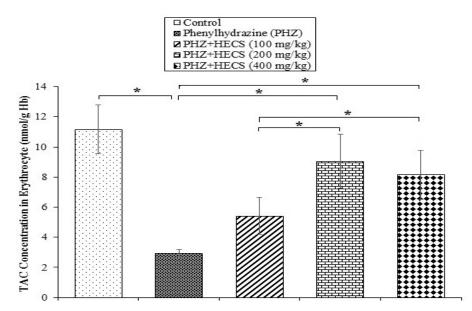


Figure 2. Effect of *Cynara scolymus* extract at different doses on TAC concentration in erythrocyte in PHZ-induced anemic rats. Data are expressed as mean±SD (n=6). * P≤ 0.05. PHZ: Phenylhydrazine; HECS: Hydroethanolic Extract of *Cynara scolymus*

Effect of *cynara scolymus* extract on MDA levels in serum and erythrocytes

A marked increase in the MDA level was found in the serum and erythrocytes of PHZ-exposed rats relative to normal rats ($P \le 0.05$). The treatment of anemic rats with increasing concentrations of Cynara scolymus extract (100-400 mg/kg) was significantly effective in decreasing the elevated level of MDA ($P \le 0.05$). No significant difference was observed in serum MDA level between HECS treated groups (P > 0.05) but a decline in efficacy of HECS in MDA concentration in RBCs was in a dose-dependent manner ($P \le 0.05$) (Figure 3 and Figure 4).

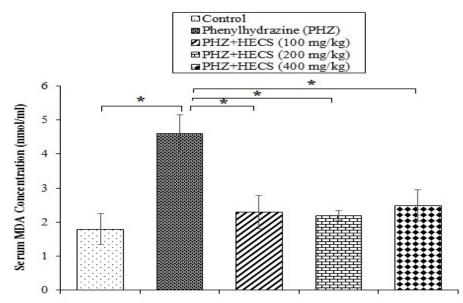


Figure 3. Effect of *Cynara scolymus* extract at different doses on serum MDA concentration in PHZ-induced anemic rats. Data are expressed as mean±SD (n=6). * *P*≤ 0.05. PHZ: Phenylhydrazine; HECS: Hydroethanolic Extract of *Cynara scolymus*

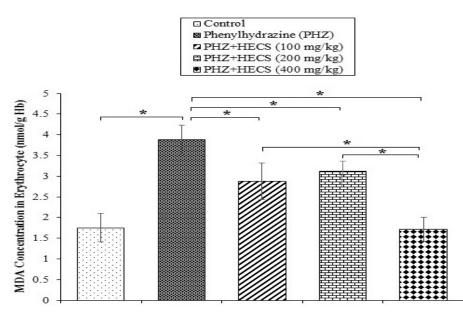


Figure 4. Effect of *Cynara scolymus* extract at different doses on MDA concentration in erythrocyte in PHZ-induced anemic rats. Data are expressed as mean±SD (n=6). * P≤ 0.05. PHZ: Phenylhydrazine; HECS: Hydroethanolic Extract of *Cynara scolymus*

Effect of *cynara scolymus* extract on SOD levels in serum and erythrocytes

The SOD activity in serum and erythrocytes has significantly decreased in PHZ group when compared with the normal control group ($P \le 0.05$). Significant increase of serum and erythrocytes SOD level was determined in rats treated with *Cynara scolymus* extract at different doses, especially at 200 mg/kg and 400 mg/kg, as compared to untreated anemic rats. Also, statistically significant difference in the SOD level was ascertained between the rats in the various groups treated with HECS ($P \le 0.05$) (Figure 5 and Figure 6).

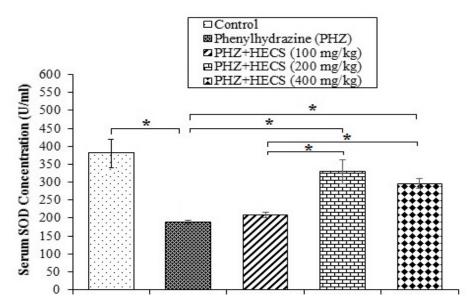


Figure 5. Effect of *Cynara scolymus* extract at different doses on serum SOD concentration in PHZ-induced anemic rats. Data are expressed as mean±SD (n=6). * P≤ 0.05. PHZ: Phenylhydrazine; HECS: Hydroethanolic Extract of *Cynara scolymus*

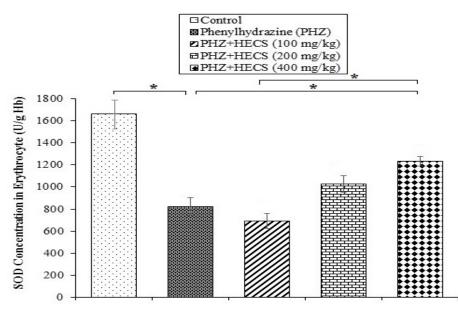


Figure 6. Effect of *Cynara scolymus* extract at different doses on SOD concentration in erythrocyte in phenylhydrazine-induced anemic rats. Data are expressed as mean±SD (n=6). * *P*≤ 0.05. PHZ: Phenylhydrazine; HECS: Hydroethanolic Extract of *Cynara scolymus*

Discussion

The purpose of the study was to verify the antioxidant effects of artichoke extract on serum and erythrocytes antioxidant capacity following hemolytic anemia induced by PHZ injection. Our results indicated that the injection of PHZ significantly decreased Hb and PCV levels in comparison with the control group. In PHZ-induced anemia, ROS-mediated damage of RBCs membrane components is thought to increase lipid peroxidation, erythrocyte membrane rigidity and fragility. In this regard, intravascular hemolysis was increased and higher levels of hemoglobin released into the serum (Pandey et al., 2014). Extract-treated animals in this study have enhanced Hb, PCV levels and RBCs numbers in comparison with PHZ group. Therefore, the beneficial impact of ethanol extract of artichoke in improving hematinic profile in PHZ-induced anemia may be due to reducing ROS which leads to establishing fragility of RBCs and restoring the total Hb concentration in the blood (Avcı et al., 2006; Rezazadeh et al., 2012, Salekzamani et al., 2019). On the other hand, it was reported that Cynara scolymus is rich in phenolic compounds belonging to different classes such as benzoic and cinnamic derivatives, flavonoids and tannins (Lattanzio et al., 2009). Salem et al. (2017) indicated that flavonoids could induce secretion of erythropoietin, which stimulates stem cells in the bone marrow to produce RBCs.

The present study demonstrated that administration of PHZ to rats results in a marked leukocytosis in comparison with control group. This leukocyte response is due to an increase in circulating mononuclear cells, primarily lymphocytes. The results of present study are in line with the results of Borley *et al.* (2010) and Xia *et al.* (2014) which indicated that leukocytosis in PHZ-treated animals occurred by increasing oxidative and nitrosative stress. The use of high doses of artichoke extract increased the number of leukocytes compared to the PHZ group, which may be due to its antioxidant capacity and its ability to stimulate the production of leucocytes and could serve as immune booster (Pérez-García *et al.*, 2000; Zapolska-Downar *et al.*, 2002).

The antioxidant capacity of whole blood is driven by the circulating RBCs that act as mobile scavengers and exert their antioxidant properties (Siems et al., 2000). If oxidative stress persists, the production of free radicals is greater than the antioxidant capacity of erythrocytes or serum. Thus, incidence of hemolytic anemia or sickle cell anemia is imminent (Chan et al., 1999). Measurement of antioxidant molecules is difficult and requires time and cost. Therefore, the total antioxidant capacity of erythrocytes can be one of the reliable indices to determining oxidative stress in erythrocytes and serum (Singh et al., 2018). Administration of PHZ causes ROS production which leads to the hemolytic anemia. There are several ways to reduce the effects of ROS on haematotoxicity, the most common of which are medicinal plants. Natural antioxidants found in foods and vegetables are one of several factors that increase the antioxidant capacity of erythrocytes (Jimenez-Escrig et al., 2003). The collection data of the present study revealed that TAC level of serum and erythrocytes in PHZ group decreased in comparison with control group, which is attributed to increased oxidative stress and ROS generation following PHZ administration (Shukla et al., 2012). Administration of Cynara scolymus extract considerably increased TAC level as compared to the PHZ group. This effect of artichoke extract might be dependent on its flavonoids. According to previous research, it was demonstrated that phenolics and flavonoids compounds have a highly antioxidant effect by reducing ROS (Petropoulos *et al.*, 2017; Sahebkar *et al.*, 2018).

Evaluation of MDA concentrations in serum and erythrocytes showed a significant increase in PHZ group compared to the control group. which in line with the results of Mozafari et al.(Mozafari *et al.*, 2016). The level of lipid peroxide is a measure of membrane damage and alterations in the structure and function of cellular membranes (Salem *et al.*, 2017).

One of the important oxidation products is MDA, which is considered as the main marker of lipid peroxidation in oxidative stress, inflammation and various health disorders (Singh et al., 2014). Recent studies have suggested that increase in MDA concentrations is one of the most important mechanisms of PHZ in development of hemolytic anemia (Gaschler et al., 2017; Gheith et al., 2018). Erythrocytes and erythrocyte membrane are more vulnerable to lipid peroxidation and oxygen radical formation because of constant exposure to high oxygen tension and richness in polyunsaturated fatty acid. Alterations in lipid composition of RBC membranes result in morphologically abnormal erythrocytes with decreased life span (Revin et al., 2016). It has been suggested that the membrane lipids play crucial roles in cellular homeostasis including the maintenance of erythrocytes structure, cellular function and viability. Increased ROS, as stated, increases lipid peroxidation and produces substances that are toxic to cells and disrupt cells structure (i.e., hemolysis) (Pandey et al., 2014; Mladenov et al., 2015). The present work showed that Cynara scolymus extract resulted in a significant reduction of the MDA concentrations in

PHZ-exposed rats. Hence, it is possible that the mechanism of this protection may be due to its antioxidant activity. In this regard, recent study has shown that *Bidens pilosa* extract with its antioxidant properties, could prevent oxidative damage and reduce MDA level in erythrocytes (Yang *et al.*, 2006).

Our results revealed that SOD activity in serum and erythrocytes has significantly decreased in PHZ group when compared with the control group. The reduction in the SOD activity may be due to the oxidation of cysteine in this enzyme by superoxide anion during its transformation to hydrogen peroxide. In other words, the decrease in SOD activity in PHZ-induced anemic rats may be due to the use of this enzyme in converting the superoxide anion $(O_2 \bullet -)$ to hydrogen peroxide (H₂O₂) (Plestina-Borjan et al., 2015). In addition, PHZ generates free radicals and ROS such as superoxide anion and hydrogen peroxide (Kamisah et al., 2014). Moreover, increased lipid peroxidation induced by PHZ in the serum and erythrocytes was accompanied by a decrease in SOD activity, which was also reported by recent studies (Maity et al., 2013; Anbara et al., 2015). Erythrocytes contain several biological defense mechanisms against free radical-induced injury, including many antioxidant enzymes such as SOD (Kolanjiappan et al., 2002). Furthermore, SOD could inhibit the accumulation of membrane proteins, which might prevent the disruption or destruction of erythrocyte wall proteins. Also, SOD activity in the cytosol of erythrocytes is about 3 times more than serum, so it could help modulate the oxidant effects of PHZ in erythrocytes. Given these points, measuring SOD can be an important criterion for the prognosis of diseases that cause hemolysis or damage to erythrocytes (Valenzuela et al., 1977; Pandey and Rizvi,

2011; Anbara *et al.*, 2018). Remarkable increments in SOD level of serum and erythrocytes were determined in anemic rats treated with *Cynara scolymus* as compared to untreated anemic rats. These positive results are consistent with another study which has demonstrated natural medicinal herbs could boost SOD activity due to presence of antioxidant compounds (Fukai *et al.*, 2011). In accordance with these findings, the study by Salem *et al.* (2017) showed that administration of ethanol extract of *C. scolymus* increased the SOD level in another oxidative stress-related disorder like diabetes, which indicates the antioxidant activity of extract.

In conclusion, PHZ caused marked oxidative stress and lipid peroxidation in rats by inhibiting the antioxidant enzymes activity. The results revealed the ameliorating effect of *Cynara scolymus* extract through declining the levels of MDA and raising the SOD and TAC activity in PHZ-induced hemolytic anemia in rats. Protective property of *Cynara scolymus* extract against PHZ-induced toxicity might be due to the presence of various phytochemical constituents with potent antioxidant activity.

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Conflict of interest

The authors declared that there is no conflict of interest.

References

- Anbara, H., Shahrooz, R., and Malekinejad, H. (2015). The protective effect of royal jelly and vitamin c coadministration on detrimental effects of phenylhydrazine-induced hemolytic anemia on the parameters of testicular tissue in adult laboratory mice. Qom Univ Med Sci J, 9(8), 1-12.
- Anbara, H., Shahrooz, R., Razi, M., Malekinejad, H., and Najafi, G. (2018). The effect of vitamin C on mice hemolytic anemia induced by phenylhydrazine: an animal model study using histological changes in testis, pre-implantation embryo development, and biochemical changes. Iran J Basic Med Sci, 21(7), 668. https:// doi.org/10.22038/IJBMS.2018.25819.6356 PMID: 30140404
- Asase, A., Kokubun, T., Grayer, R.J., Kite, G., Simmonds, M.S., Oteng-Yeboah, A.A. and Odamtten, G.T.(2008). Chemical constituents and antimicrobial activity of medicinal plants from Ghana: *Cassia sieberiana, Haematostaphis barteri, Mitragyna inermis and Pseudocedrela kotschyi*. Phytother Res. 22(8), pp.1013-1016. https://doi.org/10.1002/ptr.2392 PMID: 18618525
- Avcı, G., Kupeli, E., Eryavuz, A., Yesilada, E., and Kucukkurt, I. (2006). Antihypercholesterolaemic and antioxidant activity assessment of some plants used as remedy in Turkish folk medicine. J Ethnopharmacol, 107(3), 418-423. https://doi.org/10.1016/j.jep.2006.03.032 PMID: 16713156
- Banerjee, J., Das, A., Sinha, M. and Saha, S. (2018).
 Biological efficacy of medicinal plant extracts in preventing oxidative damage. Oxid Med Cell Longev, https://doi.org/10.1155/2018/7904349
 PMID: 30302174
- Borley, K. A., Beers, J. M., and Sidell, B. D. (2010). Phenylhydrazine-induced anemia causes nitric-oxide-mediated upregulation of the angiogenic pathway in Notothenia coriiceps. J Exp Biol., 213(16), 2865-2872. https://doi.org/10.1242/jeb.043281 PMID: 20675556
- Al-Naama, L. M., Mea'ad, K. H., and Mehdi, J. K. (2015). Association of erythrocytes antioxidant enzymes and their cofactors with markers

of oxidative stress in patients with sickle cell anemia. Qatar Med J ., 2015(2), 14. https://doi. org/10.5339/qmj.2015.14

- Chan, A. C., Chow, C. K., and Chiu, D. (1999). Interaction of antioxidants and their implication in genetic anemia. SEBM, 222(3), 274-282. https:// doi.org/10.1046/j.1525-1373.1999.d01-144.x PMID: 10601886
- Dhaliwai, G., Cornett, P.A. and Tierney, L.M. (2004). Hemolytic anemia. Am Fam Physician, 69, pp.2599-2608. PMID: 15202694
- Diallo, A., Gbeassor, M., Vovor, A., Eklu-Gadegbeku, K., Aklikokou, K., Agbonon, A., Abena, A.A., de Souza, C. and Akpagana, K. (2008). Effect of *Tectona grandis* on phenylhydrazine-induced anaemia in rats. Fitoterapia, 79(5), 332-336. https://doi.org/10.1016/j.fitote.2008.02.005 PMID: 18504074
- El-Boshy, M., Ashshi, A., Gaith, M., Qusty, N., Bokhary, T., AlTaweel, N. and Abdelhady, M. (2017). Studies on the protective effect of the artichoke (*Cynara scolymus*) leaf extract against cadmium toxicity-induced oxidative stress, hepatorenal damage, and immunosuppressive and hematological disorders in rats. Environ Sci Pollut Res Int. 24(13), pp.12372-12383. https://doi.org/10.1007/s11356-017-8876-x PMID: 28357802
- Forni, C., Facchiano, F., Bartoli, M., Pieretti, S., Facchiano, A., D'Arcangelo, D., Norelli, S., Valle, G., Nisini, R., Beninati, S. and Tabolacci, C. (2019). Beneficial role of phytochemicals on oxidative stress and age-related diseases. Biomed Res Int. https://doi.org/10.1155/2019/8748253 PMID: 31080832.
- Fukai, T., and Ushio-Fukai, M. (2011). Superoxide dismutases: role in redox signaling, vascular function, and diseases. Antioxid Redox Signal, 15(6), 1583-1606. https://doi.org/10.1089/ ars.2011.3999 PMID: 21473702
- Gaschler, M. M., and Stockwell, B. R. (2017). Lipid peroxidation in cell death. Biochem Biophys Res Commun, 482(3), 419-425. https://doi. org/10.1016/j.bbrc.2016.10.086 PMID: 28212725
- Gebhardt, R. and Fausel, M. (1997). Antioxidant and hepatoprotective effects of artichoke extracts and constituents in cultured rat hepato-

cytes. Toxicol In Vitro. 11(5), pp.669-672. https://doi.org/10.1016/s0887-2333(97)00078-7 PMID: 20654368

- Gheith, I., and El-Mahmoudy, A. (2018). Laboratory evidence for the hematopoietic potential of *Beta vulgar*is leaf and stalk extract in a phenylhydrazine model of anemia. Braz J Med Biol Res, 51(11). https://doi.org/10.1590/1414-431X20187722 PMID: 30328935
- Jimenez-Escrig, A., Dragsted, L. O., Daneshvar, B., Pulido, R., and Saura-Calixto, F. (2003). In vitro antioxidant activities of edible artichoke (*Cy-nara scolymus* L.) and effect on biomarkers of antioxidants in rats. J Agric Food Chem, 51(18), 5540-5545. https://doi.org/10.1021/jf030047e
- Kamisah, Y., Lim, J.J., Lim, C.L. and Asmadi, A.Y. (2014). Inhibitory effects of palm tocotrienol-rich fraction supplementation on bilirubin-metabolizing enzymes in hyperbilirubinemic adult rats. PloS one, 9(2), p.e89248. https:// dx.doi.org/10.1371%2Fjournal.pone.0089248 PMID: 24586630
- Kaur, R., Ghanghas, P., Rastogi, P., and Kaushal, N. (2019). Protective role of selenium against hemolytic anemia is mediated through redox modulation. Biol Trace Elem Res, 189(2), 490-500. https://doi.org/10.1007/s12011-018-1483-y PMID: 30151563
- Kolanjiappan, K., Manoharan, S. and Kayalvizhi, M. (2002). Measurement of erythrocyte lipids, lipid peroxidation, antioxidants and osmotic fragility in cervical cancer patients. Clin Chim Acta. 326(1-2), pp.143-149. https://doi.org/10.1016/ s0009-8981(02)00300-5 PMID: 12417105
- Lattanzio, V., Kroon, P.A., Linsalata, V. and Cardinali, A. (2009). Globe artichoke: a functional food and source of nutraceutical ingredients. J Funct Foods, 1(2), pp.131-144. https://doi. org/10.1016/j.jff.2009.01.002
- Lee, S.H., Suh, H.J., Lee, H.S., Park, Y., Park, J.W. and Jung, E.Y. (2012). Hematopoietic effect of *Bacillus subtilis*–fermented antler extract on phenylhydrazine-induced hemolytic anemia in Sprague–Dawley rats. J Med Food, 15(9), pp.774-780. https://doi.org/10.1089/ jmf.2012.2264 PMID: 22870931

Luangaram, S., Kukongviriyapan, U., Pakdeechote,

P., Kukongviriyapan, V., and Pannangpetch, P. (2007). Protective effects of quercetin against phenylhydrazine-induced vascular dysfunction and oxidative stress in rats. Food Chem Toxicol, 45(3), 448-455. https://doi.org/10.1016/j. fct.2006.09.008 PMID: 17084956

- Maity, S., Nag, N., Chatterjee, S., Adhikari, S. and Mazumder, S. (2013). Bilirubin clearance and antioxidant activities of ethanol extract of *Phyllanthus amarus* root in phenylhydrazine-induced neonatal jaundice in mice. J Physiol Biochem, 69(3), pp.467-476. https://doi.org/10.1007/ s13105-013-0234-y PMID: 23318962
- Mladenov, M., Gokik, M., Hadzi-Petrushev, N., Gjorgoski, I., and Jankulovski, N. (2015). The relationship between antioxidant enzymes and lipid peroxidation in senescent rat erythrocytes. Physiol Res, 64(6), 891. PMID: 26047376
- Mozafari, A. A., Shahrooz, R., Ahmadi, A., Malekinjad, H., and Mardani, K. (2016). Protective effect of ethyl pyruvate on mice sperm parameters in phenylhydrazine induced hemolytic anemia. Vet Res Forum, 7(1), 63. PMID: 27226889
- Pandey, K., Meena, A. K., Jain, A., and Singh, R. K. (2014). Molecular mechanism of phenylhydrazineinduced haematotoxicity: A review. Ame J Phytomed Clin Therapeut, 2, 390-394.
- Pandey, K. B., and Rizvi, S. I. (2011). Biomarkers of oxidative stress in red blood cells. Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub., 155(2). PMID: 21804621
- Paul, S., Naaz, S., Ghosh, A. K., Mishra, S., Chattopadhyay, A., and Bandyopadhyay, D. (2018). Melatonin chelates iron and binds directly with phenylhydrazine to provide protection against phenylhydrazine induced oxidative damage in red blood cells along with its antioxidant mechanisms: an in vitro study. Melatonin Res, 1(1), 1-20. https://doi.org/https://doi.org/10.32794/mr11250009.
- Pérez-García, F., Adzet, T., and Cañigueral, S. (2000). Activity of artichoke leaf extract on reactive oxygen species in human leukocytes. Free Radic Res., 33(5), 661-665. https://doi.org/10.1080/10715760000301171
 PMID: 11200096
- Petropoulos, S. A., Pereira, C., Barros, L., and Ferreira, I. C. (2017). Leaf parts from Greek artichoke genotypes as a good source of bioactive

compounds and antioxidants. Food Funct, 8(5), 2022-2029. https://doi.org/10.1039/c7fo00356k PMID: 28492621

- Plestina-Borjan, I., Katusic, D., Medvidovic-Grubisic, M., Supe-Domic, D., Bucan, K., Tandara, L., and Rogosic, V. (2015). Association of age-related macular degeneration with erythrocyte antioxidant enzymes activity and serum total antioxidant status. Oxid Med Cell Longev., 2015. https://doi.org/10.1155/2015/804054 PMID: 25815109
- Prasad, S. Y., Hari, P., Shajina, M., Mirshad, P. V., and Rahiman, F. O. (2018). Hematinic and antioxidant potential of aqueous extract of *Sesamum indicum* seeds against phenylhydrazine-induced hemolytic anemia in albino rats. Natl J Physiol Pharm., 8(8), 1092-1096. https://doi. org/10.5455/njppp.2018.8.0310831032018
- Revin, V.V., Gromova, N.V., Revina, E.S., Martynova, M.I., Seikina, A.I., Revina, N.V., Imarova, O.G., Solomadin, I.N., Tychkov, A.Y. and Zhelev, N. (2016). Role of membrane lipids in the regulation of erythrocytic oxygen-transport function in cardiovascular diseases. Biomed Res Int. https://doi.org/10.1155/2016/3429604 PMID: 27872848
- Rezazadeh, A., Ghasemnezhad, A., Barani, M., and Telmadarrehei, T. (2012). Effect of salinity on phenolic composition and antioxidant activity of artichoke (*Cynara scolymus* L.) leaves. Res J Med Plant, 6, 245-252. http://dx.doi. org/10.3923/rjmp.2012.245.252
- Sahebkar, A., Pirro, M., Banach, M., Mikhailidis, D. P., Atkin, S. L., and Cicero, A. F. (2018). Lipid-lowering activity of artichoke extracts: a systematic review and meta-analysis. Crit Rev Food Sci Nutr, 58(15), 2549-2556. https://doi.org/10.1 080/10408398.2017.1332572 PMID: 28609140
- Salekzamani, S., Ebrahimi-Mameghani, M., and Rezazadeh, K. (2019). The antioxidant activity of artichoke (*Cynara scolymus*): A systematic review and meta-analysis of animal studies. Phytother Res., 33(1), 55-71. https://doi. org/10.1002/ptr.6213 PMID: 30345589
- Salem, M. B., Affes, H., Ksouda, K., Dhouibi, R., Sahnoun, Z., Hammami, S., and Zeghal, K. M. (2015). Pharmacological studies of artichoke leaf extract and their health benefits. Plant Foods Hum

Nutr, 70(4), 441-453. https://doi.org/10.1007/ s11130-015-0503-8 PMID: 26310198

- Salem, M.B., Kolsi, R.B.A., Dhouibi, R., Ksouda, K., Charfi, S., Yaich, M., Hammami, S., Sahnoun, Z., Zeghal, K.M., Jamoussi, K. and Affes, H. (2017). Protective effects of *Cynara scolymus* leaves extract on metabolic disorders and oxidative stress in alloxan-diabetic rats. BMC Complement Altern Med, 17(1), p.328. https://dx.doi.org/10.1186%2Fs12906-017-1835-8 PMID: 28629341
- Shetlar, M.D. and Hill, H.A. (1985). Reactions of hemoglobin with phenylhydrazine: a review of selected aspects. Environ Health Perspect. 64, pp.265-281. https://doi.org/10.1289/ ehp.8564265 PMID: 3007094.
- Shimoda, H., Ninomiya, K., Nishida, N., Yoshino, T., Morikawa, T., Matsuda, H., and Yoshikawa, M. (2003). Anti-hyperlipidemic sesquiterpenes and new sesquiterpene glycosides from the leaves of artichoke (*Cynara scolymus* L.): structure requirement and mode of action. Bioorg Med Chem Lett., 13(2), 223-228. https://doi.org/10.1016/ s0960-894x(02)00889-2 PMID: 12482428
- Shukla, P., Yadav, N.K., Singh, P., Bansode, F.W. and Singh, R.K. (2012). Phenylhydrazine induced toxicity: a review on its haematotoxicity. Int J Basic Appl Med Sci, 2(2), pp.86-91.
- Siems, W. G., Sommerburg, O., and Grune, T. (2000). Erythrocyte free radical and energy metabolism. Clin Nephrol., 53(1 Suppl), S9-17. PMID:10746800
- Singh, Z., Karthigesu, I. P., Singh, P., and Rupinder, K. A. U. R. (2014). Use of malondialdehyde as a biomarker for assessing oxidative stress in different disease pathologies: a review. Iran J Public Health, 43(Supple 3), 7-16.
- Singh, A. K., Singh, S., Garg, G., and Rizvi, S. I.

(2018). Rapamycin mitigates erythrocyte membrane transport functions and oxidative stress during aging in rats. Arch Physiol Biochem, 124(1), 45-53. https://doi.org/10.1080/1381345 5.2017.1359629 PMID: 28758804

- Sung, Y.M., Gayam, S.R. and Wu, S.P. (2013). The oxidation of phenylhydrazine by tyrosinase. Appl Biochem Biotechnol. 169(8), pp.2420-2429. https://doi. org/10.1007/s12010-013-0165-7 PMID: 23456281
- Valenzuela, A., Rios, H., and Neiman, G. (1977). Evidence that superoxide radicals are involved in the hemolytic mechanism of phenylhydrazine. Experientia, 33(7), 962-963. https://doi. org/10.1007/BF01951306 PMID: 196888
- Xia, N., Pautz, A., Wollscheid, U., Reifenberg, G., Förstermann, U., and Li, H. (2014). Artichoke, cynarin and cyanidin downregulate the expression of inducible nitric oxide synthase in human coronary smooth muscle cells. Molecules, 19 (3), 3654-3668. https://doi.org/10.3390/molecules19033654 PMID: 24662080
- Yang, H.L., Chen, S.C., Chang, N.W., Chang, J.M., Lee, M.L., Tsai, P.C., Fu, H.H., Kao, W.W., Chiang, H.C., Wang, H.H. and Hseu, Y.C. (2006). Protection from oxidative damage using *Bidens pilosa* extracts in normal human erythrocytes. Food Chem Toxicol., 44(9), 1513-1521. https://doi.org/10.1016/j.fct.2006.04.006 PMID: 16765500
- Zapolska-Downar, D., Zapolski-Downar, A., Naruszewicz, M., Siennicka, A., Krasnodębska, B. and Kołodziej, B. (2002). Protective properties of artichoke (*Cynara scolymus*) against oxidative stress induced in cultured endothelial cells and monocytes. Life Sci. 71(24), pp.2897-2908. https://doi.org/10.1016/s0024-3205(02)02136-7 PMID: 12377270

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بهبود وضعیت پراکسیداسیون لیپید و آنزیمهای آنتی اکسیدانی در سرم و گلبولهای قرمز رتهای نر مبتلا به آنمی القاء شده توسط فنیل هیدرازین: نقش محافظتي عصاره كنكر فرنكي

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*چکید*ه

زمینه مطالعه: آنمی همولیتیک اختلالی است که با تخریب زودرس گلبولهای قرمز مشخص می شود. فنیل هیدرازین سبب القاء استرس اکسیداتیو و تشکیل گونههای واکنش پذیر اکسیژن شده که منجر به بروز آنمی همولیتیک می شود. کنگر فرنگی بدلیل تر کیبات آنتی – کسیدانی خود برای اهداف مختلف درمانی در طب سنتی استفاده شده است.

هدف: مطالعه حاضر جهت بررسی اثرات عصاره کنگر فرنگی بر آنمی القاء شده توسط فنیل هیدرازین در رتهای نر طراحی شد.

روش کار: آنمی همولیتیک با تزریق داخل صفاقی فنیل هیدرازین (۴۰ mg/kg) به مدت ۲ روز القاء شد. ۳۰ سر رت نر نژاد ویستار به پنج گروه تقسیم شدند. گروه اول (کنترل نرمال). گروه ۲ فقط فنیل هیدرازین را دریافت کرد (کنترل آنمیک). گروه های ۳ تا ۵ با کنگر فرنگی به ترتیب با دوزهای ۲۰۰، ۲۰۰ و ۴۰۰ میلی گرم به ازای کیلوگرم وزن بدن از روز ۲ تا ۱۵ پس از تزریق فنیل-هیدرازین گاواژ شدند. در پایان دوره درمانی، نمونههای خون جهت ارزیابی پارامترهای هماتولوژی، سطح مالون دی آلدئید (MDA) و فعالیت آنزیمهای آنتی اکسیدانی مانند سوپراکسید دیسموتاز (SOD) و ظرفیت آنتی اکسیدانی تام (TAC) در سرم و گلبول های قرمز جمع آوری شد.

نتاییج: در رتهای کم خون، سطح MDA سرم و گلبولهای قرمز افزایش یافت، اما فعالیت SOD و TAC در مقایسه با گروه کنترل بطور معنیداری کاهش یافت (۲۰۵۵ ≥ P. این تغییرات با درمان توسط کنگر فرنگی در دوزهای مختلف بهبود یافت (۲۰۵۵ ≤ P). همچنین، چندین پارامتر هماتولوژیکی بهبود یافته پس از تجویز کنگر فرنگی در رتهای کم خون مشاهده شد (۲۰۵۵).

نتیجه گیری نهایی: عصاره کنگر فرنگی دارای خاصیت محافظتی در برابر استرس اکسیداتیو ناشی از فنیل هیدرازین بوده که احتمالاً به دلیل فعالیت آنتیاکسیدانی عصاره این گیاه است.

واژەھايكليدى:

استرس اكسيداتيو، أنمى هموليتيك، رت، كنكر فرنكى، فنيل هيدرازين.