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6 **Prevalence of *Paramphistomum* and its associated factors in Cattle**
7 **slaughtered at Boko slaughter house, Fedis District, Eastern Hararghe**
8 **Zone, Ethiopia**

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12 Ethiopia

13
14 **Abstract**

15 **BACKGROUND:** Bovine paramphistomosis is one of the most important parasitic
16 diseases of cattle causing mortality and production losses in various parts of Ethiopia.

17 **OBJECTIVES:** A cross sectional study was carried out in Fedis district from April 2019
18 to October 2020 to determine the prevalence of *Paramphistomum* and its associated
19 factors in cattle slaughtered at Boko slaughter house.

20 **METHODS:** A total of 384 slaughtered cattle were selected by systematic random
21 sampling and post-mortem examination of rumen and reticulum was conducted to check
22 the presence or absence of adult *Paramphistomum*. The parasite was examined
23 macroscopically and microscopically to appreciate the morphology of adult flukes.

24 **RESULTS:** Of 384 examined cattle, the prevalence of *Paramphistomum* was recorded
25 in 156(40.6%). Of the total 156 positive samples 52.34%, 33.06% and 14.6% were
26 infected with *Paramphistomum clavula*, *paramphistomum cervi* and mixed infection
27 respectively. In this study, highest rate of infection was recorded in wet season. Among
28 assessed factors; body condition and origin of animals was significant variation ($P <$
29 0.05) with infection.

30 **CONCLUSIONS:** This finding indicated the highest infection of *paramphistomum* was
31 recorded cattle. Therefore, the farmers should be practiced mass de-worming of animals
32 using selected anthelmintic therapy for flukes.

33 **Keywords:** Abattoir, Boko, cattle, *Paramphistomum*, Risk Factors.

34 **Introduction**

35 Ethiopia is believed to have the largest livestock population in Africa, yet produces
36 insufficient animal protein and other livestock products to meet the demand of human
37 population. The livestock sector has been contributing a considerable portion to the
38 economy of the country and still promising to rally around the economic development of
39 the country. However, production and productivity are low in Ethiopia, due to poor
40 nutrition, reproduction insufficiency, poor breed improvement, management constraints,
41 and prevailing of diseases (Ayele *et al.*, 2003). Among diseases, parasitism is the major

42 problems that affect the productivity of livestock worldwide. Amphistomosis is globally
43 distributed, but the highest prevalence has been reported from tropical and sub-tropical
44 regions, particularly Africa, Asia, Australia, Eastern Europe and Russia. It is caused by
45 digenetic flukes belong to the family Paramphistomidae. The life cycle of these
46 Trematodes involves snail as an intermediate host (Huson *et al.*, 2017).

47 The epidemiology of *Paramphistomum* infection in cattle is determined by several factors
48 governed by parasite-host-environment interactions (Martinez-Ibeas *et al.*, 2016). It is
49 also influenced by the climatic requirement for egg hatching, development and survival
50 of the larvae in pasture (Ozidal *et al.*, 2010). Adults *Paramphistomum* are found in the
51 rumen and reticulum whilst immature parasites are found in the duodenum. Adult
52 *Paramphistomum* flukes parasitize mainly in the fore stomachs of cattle causing irregular
53 rumination, lower nutrition conversion, loss of body condition, decrease milk production,
54 and reduction of fertility rate (Getenet *et al.*, 2016). Severe infections with massive
55 number of immature parasites migrating through the intestinal tract cause acute parasitic
56 gastroenteritis with high morbidity and mortality rates, particularly in young animals
57 (Maitra *et al.*, 2014; Huson *et al.*, 2017).

58 *In spite of the aforementioned prevailing situation and the presence of a number of*
59 *problems due to gastrointestinal parasites there is scarcity of well-documented*
60 *information on the occurrence of Paramphistomum in ruminants in Ethiopia and also*
61 *there is no any conducted research in Fedis district. knowledge of the prevalence and*
62 *risk factors would help in implementing of strategies and policies for control and*
63 *prevention of bovine Paramphistomosis in the study area. Therefore, the objectives of this*

64 *study was to determine the prevalence and associated factors of Paramphistomum in*
65 *cattle slaughtered in the study area.*

66 **Materials and Methods**

67 **Study Area**

68 The study was carried out in Fedis district Boko slaughter house. The district is found in
69 Eastern Hararghe, Oromia Regional State, Ethiopia, at a geographical coordinate of 8°
70 49' 43.3"N latitude and 42° 0' 45.57"E longitude and an elevation of 1285m above sea
71 level. It is divided into three major agro-climatic zones, namely highland, mid-highland
72 and lowland. The district consists of two seasons, mainly wet season from April to
73 September, and the dry season from October to March. The main occupation of the
74 population in these rural Kebeles are mixed farming system. Livestock species include:
75 cattle, sheep, goat, donkey and Poultry. The livelihood of the population is 93.8% agro-
76 pastoralist while the rest, 6.2% are urban dwellers (FDOA, 2020).

77 **Study Animals**

78 The study animals were cattle (local breeds) of different sexes, ages groups and body
79 conditions brought from highland, mid highland and lowland areas to the slaughter house.
80 The body condition of animals was carried out through observation and palpation on the
81 body fat deposits under the skin around the base of the tail, spine, and hip. Accordingly it
82 was categorized in three namely good, medium and poor. Ageing of the cattle was based
83 on rostral dentition as described by Lasisi *et al.* (2002). Cattle aged (< 3 years old) were
84 classified as young while (> 3 years old) were considered as adults. Sexual differentiation
85 was based on the appearance of external genitals as described by Yunusa *et al.* (2013).

86 **Study Design and Sample Size Determination**

87 A cross sectional study was conducted in Fedis District from April 2019 to October 2020
88 to determine the prevalence and associated factors of *Paramphistomum* in cattle
89 slaughtered at Boko slaughter house using post-mortem examination. The number of
90 cattle required for the study was calculated based on the formula given by (Thrusfield and
91 Christley, 2018). The sample size was determined based on expected prevalence of 50%,
92 confidence interval of 95% and desired level of precision of 5%.

$$93 n = 1.96^2 [P_{exp} (1 - P_{exp})] / d^2$$

94 Where, n = required sample size, P_{exp} = expected prevalence, d = desired absolute
95 precision. Therefore, based on the above formula 384 cattle slaughtered at Boko
96 slaughter house were selected for this study.

97 **Sampling Methods**

98 The samples were collected from the slaughter house at the time of slaughtering process
99 was conducted. Systematic random sampling technique was used to select the study
100 animals i.e. the first animal was taken randomly and the next animals were selected on an
101 interval of three.

102 **Study Methodology**

103 During ante mortem inspection, my own identification was carried out using marker on
104 the head for each of study animals. General physical examinations of animals were
105 conducted and details about sex, age, origin and body condition of the animals was
106 recorded. During post-mortem examination, *rumen* and *reticulum* of the selected animals

107 was systematically inspected to check the presence of adult *Paramphistomum*. If the
108 evidence of *paramphistomum* were found they are recorded separately as positive. Then
109 secondary examination was conducted by further incisions of the rumen and reticulum.
110 The parasite was examined macroscopically and microscopically to appreciate the
111 morphology of adult flukes. Final identification of *Paramphistomum* was done based on
112 morphology of flukes; shape, posterior sucker (acetabulum), anterior sucker, terminal
113 genitalium and tegumental papillae following the standard guidelines (Urquhart *et al.*,
114 1996).

115 **Data Management and Analysis**

116 All of the data collected from each of slaughtered animals were entered into Microsoft
117 Excels spread sheet and analyzed with SPSS Version 20. Chi- square test (X^2) was used
118 to show the variation between different risk factors such as sex, age, body condition and
119 origin of animals. ($P < 0.05$) was considered to be statistically significant.

120 **Results**

121 Out of the total of 384 examined animals, an overall prevalence of *Paramphistomum* was
122 156 (40.6%). This finding indicated that statistically significant variation ($P < 0.05$)
123 between with the prevalence of 52.6% and 32.7% of *Paramphistomum clavula* and
124 *Paramphistomum cervi* respectively (Table 2). The current finding showed that the
125 infection was almost similar in both species of animals. However it shows slightly higher
126 in old than adult animals, but there was no statistical significant difference ($P > 0.05$) in
127 relation to sexes and age groups of animals (Table 3). This finding showed that,
128 statistically significant variation ($P < 0.05$) with respect to origin the animals with highest

129 prevalence of *Paramphistomum* was recorded in cattle brought from highland than mid
130 highland and lowland areas (Table 4). This finding showed that, statistically significant
131 variation ($P < 0.05$) with respect to body condition of the animals with highest infection
132 was recorded in poor body conditioned animals followed by medium and good body
133 conditions (Table 5).

134 Discussion

135 Out of 384 selected animals, the overall prevalence of *Paramphistomum* was 40.6% that
136 was in line with the findings of Melaku and Addis (2012) and Gudeta and Biyansa (2019)
137 in Ethiopia and Modgy *et al.* (2009) in Egypt they reported 41.5%, 40.1% and 38.92%
138 respectively. However, the current study found a higher rate of infection than studies
139 conducted in Turkey, Ethiopia and Germany by Ozdal *et al.* (2010), Tagesse *et al.* (2014)
140 and Forstmaier *et al.* (2021) who reported 8.95% and 6.7% respectively. This finding was
141 slightly lower than 65.7% reported by Tsegabirhan *et al.* (2015) in Ethiopia and 91.16%
142 reported by Chowdhury *et al.* (2019) in Bangladesh. This difference might be due to
143 difference in sample size, season of the study, management system and environmental
144 conditions.

145 In this study finding, two most common species of *paramphistomum* were identified the
146 with the prevalence of 52.34% and 33.06% were *Paramphistomum clavula* and
147 *Paramphistomum cervi* respectively, whereas 14.6% of animals were positive for mixed
148 infection of *paramphistomum* species. This finding was agrees with the findings of
149 Ayalew *et al.* (2016) who reported the highest infection rate of *paramphistomum clavula*
150 than other species. The highest infection rate of cattle with *Paramphistomum* spp. was

151 observed in August and gradually decreased in October. This finding was agrees with the
152 previous finding by Nayab *et al.* (2017) who reported that prevalence of
153 *Paramphistomum* spp. was highest in cattle in March and the lowest in November. It has
154 been described that the bionomic requirements for breeding of *Planorbis* snails and
155 development of intramolluscan stages of flukes often reach the optimum threshold during
156 the wet months (Radostits *et al.*, 2000).

157 Sex and age groups was not significant variation ($P > 0.05$) in relation to infection rate.
158 This finding is in agreement with other findings in Iran and in Ethiopia (Javad *et al.*,
159 2015; Gudeta and Biyansa, 2019) they reported insignificant difference between sexes
160 and age groups cattle and also similar to the finding from Bangladesh by Sabbir Ahmad
161 *et al.*(2020) and Azoro (2021) who reported the prevalence of gastrointestinal parasites
162 was not significant difference in relation to sexes of animals. This is due to the fact that
163 all ages and both sexes have the same chance to ingest the infective stage larvae during
164 grazing and equally susceptible to infection.

165 This finding reported that the highest infection rate was recorded in poor body
166 conditioned animals than medium and good. There is significant difference ($P < 0.05$) in
167 relation to body conditions was observed. In agreement with this finding, Tagesse *et al.*
168 (2014) and Tsegabirhan *et al.* (2015) in Ethiopia reported significant variation ($P < 0.05$)
169 between prevalence of *Paramphistomum* and body conditions. The difference of result
170 may be due to the difference in immunity of the host and the fact that cattle with good
171 body condition expected to be de-wormed and are more able to resist infection from
172 others. It was similarly observed among the few cases which we encountered with heavy

173 fluke infestation that the animals appeared markedly emaciated with poor body
174 conditions.

175 The current study finding showed that, the highest prevalence of *Paramphistomosis* was
176 recorded in cattle brought from highland than mid highland and lowland areas of origin.
177 There was statistically significant variation ($P < 0.05$) between origin of animals. This
178 finding is agrees with Tsegabirhan *et al.* (2015) from Ethiopia who reported significant
179 difference in relation to origin of animals. But it is disagreed with Getenet *et al.* (2016) in
180 Ethiopia who reported insignificant variation between origin of animals. These
181 differences might be due to the differences in sample size, diagnostic technique, climate,
182 ecology, and livestock management system (Javad *et al.*, 2015). In General, this study
183 revealed that the highest prevalence of *paramphistomum* infection was recorded in cattle
184 in study area and based on only slaughtered animals may have some limitation on
185 results.

186 **Conclusion**

187 In this study, *Paramphistomum* was found to be prevalent in cattle. This will be a
188 hindrance to the livestock production by causing remarkable direct or indirect losses of
189 livestock productions and productivity in the study area. In the present study an overall
190 prevalence of 40.6% were infected by adult *paramphistomum*. Age, sex, body condition
191 scores and origin of animals were assessed as risk factors for occurrence of *bovine*
192 *paramphistomum*. Among the risk factors, body conditions and origin of animals was
193 statistically significant variation ($P < 0.05$) with infection rate of *paramphistomum*, but

194 sexes and age groups was not significant difference. Therefore, based on the above
195 conclusions the following recommendations were forwarded.

- 196 • Awareness creation for owners to improve feeds provision in order to obtain
197 good body condition to provide sufficient level of resistance against infections.
- 198 • Integrated strategies should be implemented for control and prevention of
199 infection using mass de-worming animals by selected anthelmintic.
- 200 • Further detail epidemiological and seasonal study should be carried out, so as to
201 design appropriate control strategies on the area.

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203 • **Conflicts of Interest**

- 204 • The Author have not declared any conflict of interest.

205 • **Acknowledgments**

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208 slaughter house worker's to allow sampling to accomplish this study.

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293 indigenus sheep using multifactorial discriminant analysis. International Journal of
294 Biodiversity Conservation 5: 661–665.
- 295 • Table 1. Overall prevalence of *bovine paramphistomum*

| N_o of Cattle Examined | N_o of Positive | Prevalence (%) | 95% CI |
|-----------------------------------------|----------------------------------|-----------------------|---------------|
| 384 | 156 | 40.6% | [35.6 – 45.6] |

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300 Table 2: Prevalence of *bovine Paramphistomum* based on species distribution

| Species | N_o of Positive | Prevalence (%) | X² (P-value) |
|-------------------------------|----------------------------------|-----------------------|--------------------------------|
| <i>Paramphistomum clavula</i> | 82 | 52.6% | |
| <i>Paramphistomum cervi</i> | 51 | 32.7% | |
| Mixed Infection | 23 | 14.7% | |

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308 Table 3: Prevalence of *bovine paramphistomum* based on sex and age groups

| Risk Factors | Variables | N_o of Examined | N_o of Positive (%) | X² (P-Value) |
|---------------------|------------------|----------------------------------|--------------------------------------|--------------------------------|
|---------------------|------------------|----------------------------------|--------------------------------------|--------------------------------|

| | | | | |
|-------------------|--------------|-----|------------|--------------|
| Sexes | Male | 291 | 117(40.2%) | 0.06(0.146) |
| | Female | 93 | 39(42.1%) | |
| Age Groups | Adult | 284 | 109(38.4%) | 2.014(>0.05) |
| | Old | 100 | 47(47.2%) | |
| | Total | 384 | 156(40.6%) | |

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316 Table 4: Prevalence of *bovine paramphistomum* based on body conditions of animals

| Body conditions | No of Examined | No of Positive (%) | X²(P-value) |
|------------------------|-----------------------|---------------------------|-------------------------------|
| Good | 90 | 30(33.3%) | 11.923(0.01) |
| Medium | 200 | 74(37.2%) | |
| Poor | 94 | 52(55.3%) | |
| Total | 384 | 156(40.6%) | |

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322 Table 5: Prevalence of *bovine paramphistomosis* based on origin of animals.

| Origin of Animals | N_o of Examined | N_o of Positive (%) | X²(P-value) |
|--------------------------|----------------------------------|--------------------------------------|-------------------------------|
| Highland | 140 | 71(50.7%) | 12.853(0.005) |
| Midhighland | 140 | 56(40%) | |
| Lowland | 104 | 29(28%) | |
| Total | 384 | 156(40.6%) | |

Uncorrected Proof