

Comparative Study of Bacterial Contamination between Local Iraqi Sheep and Goat Semen

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Abstracts

Background: Semen contamination is a key factor in decreasing fertility and the seasonal effect may act in this part.

Objectives: This study was designed to detect semen contamination in ovine and caprine during different seasons.

Methods: Six fully mature male of both sheep and goat were subjected to electro-ejaculator collection twice monthly for the period 1/2/2022 to 31/1/2023, (Spring, 1/2/2022-30/4/2022,

20 Summer, 1/5/2022-31/7/2022, Autumn 1/8/2022-31/10/2022, Winter 1/11/2022-31/1/2023), for
studying the seasonal effect. A total of 288 semen samples were collected from both species (36
samples from each per season). Bacterial isolation and identification applied to all samples.
Results: The results indicated that sheep semen had four different types of bacterial
contamination with higher number in contaminated samples than goat which showed five
different bacterial isolation, with no significant differences ($P>0.05$) in bacterial contamination
25 under effect of different seasons. The result of the herein study indicated that the sheep had four
different types of contaminated bacteria with higher contamination number than goat, while the
goat show five different bacterial isolation. There was no significant differences ($P>0.05$) in
bacterial contamination under effect of different seasons.

Conclusions: from this we conclude that the different seasons appears to have no effect on
30 bacterial contamination of semen in sheep and goat, meanwhile the isolation of all bacteria types
was considered as normal flora in both small ruminants' species.

Keyword: sheep, goat, semen, bacterial contamination, seasonal.

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Introduction

Microorganisms can affect male reproductive function either directly (reduce sperm motility, acrosomal reaction reduction or increase sperm deformity) or indirectly (reactive oxygen production) (Enwuru *et al.*, 2016). However, disease caused by them may reduce reproductive system immunity (Noakes *et al.*, 2019). Also, mating, or polluted semen, is considered one of the factors of uterus infection (Pohjanvirta *et al.*, 2020), so the male is responsible for semen pollution through contamination during semen collection with glans penis and prepuce (Zaid and Al-Zubaidy, 2009). Polluted semen negatively affects fertilization and acts as a genital disease carrier (Zaid and Al-Zubaidy, 2009). Semen contaminated with pathogens increases the threat to decrease fertility and reproduction effectiveness (Russell *et al.*, 1997). Every semen ejaculates may contain contaminations of some nonpathogenic microbes which were not used in artificial insemination procedures. The massive gathering of microbes leads in infertile mating (Thacker *et al.*, 1984). Semen contamination with either pathogenic or non-pathogenic bacteria occur during semen processing and semen storage, then these microbes will be transported through the semen into genital tract and cause serious diseases due to bacteremia or/and viraemia infections (Thibier and Guerin, 2000). Many studies were identifying bacterial contamination in the frozen semen of farm animals such as *E. coli*, *Staph. aureus*, and *Pseudo. aeruginosa* (Thibier and Guerin, 2000; and Hobson *et al.*, 2013). Semen that contains variety of microorganisms reduces the survival rate and fertility of sperms; resulting decrease in offspring, so that the suppression contamination with microbes in semen is essential in artificial insemination of the breeding stock (Schulze *et al.*, 2020). Semen quality decreased when semen samples are contaminated with bacteria, fungi and viruses, these pathogens transmitted to next generation (Mitra *et al.*, 2016). Reproductive disorder may happen due to contamination with specific microbial agents during semen collection, handling and preservation (Sannat *et al.*, 2015). There is a correlation between bacterial load and semen quality as it had had been shown that the increase of bacterial numbers led to declivity in sperm motility and viability (Reda *et al.*, 2020). Meena *et al.*, (2015 and 2017),

Reported that bulls frozen semen with higher microbial numbers are of highly negative
65 significant correlation ($P < 0.01$) with progressive motility and, vitality of sperms. Also, microbial
toxins can indirectly affect sperms motility (Wang *et al.*, 2021). Boar semen contaminated with
bacteria was affiliated with a decline in sperm viability (Luther *et al.*, 2023). Bacterial presence
inside semen results in nutrients competition and metabolic byproduction that harm sperms
(Luther *et al.*, 2023), in addition to all of that, some bacteria, when they die, cause damage to
70 spermatozoa due to the release of lipopolysaccharides contained inside their walls. Bacterial
Inflammation of female genitalia may occur after being inseminated with infected semen
(Morrell and Wallgren, 2014). As above the bacterial contamination of semen cause adverse
effect on sperm quality, either directly, on nutrients supply for sperms which is represented in
semen diluents, or indirectly by formation of endotoxins and metabolic toxic byproducts
75 (Fraczek and Kurpisz, 2015). Some of studies were conducted in order to study the bacterial
contamination (Hanoun and Al-Samrrae, 2019; Al-Taii and Yousif, 2019, Al-Taee *et al.*, 2019;
Razook *et al.*, 2020; Gharban and Yousif, 2020; Foroutan *et al.*, 2022; Gaddafi *et al.*, 2023 and
Anvar *et al.*, 2023) without dealing with semen samples. While other studies address the season
effect on genital organs without refereeing to bacterial contaminations (Ibrahim and Zaid, 2015;
80 Zaid, 2017; Ibahim and Zaid, 2017a; 2017b), one study deals with breeding season effect
(Karasahin *et al.*, 2023). This study was designed to investigate the microbial contamination
types among sheep and goat in 4 consecutive seasons in Iraq for the first time.

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Material and Methods

90 **Animals:** The study was conducted in a private property in Baghdad province from 1/2/2022 to 31/1/2023. Animals were kept in ideal condition of management during the course of the study. Semen collected by electro-ejaculator every 15 days from six mature (3 years old) rams and bucks that were used for breeding during the course of the study.

To study the seasonal effect, a total collection of 288 ejaculates along the year were spread into:

- 1) Spring season (2/2022, 3/2022 and 4/2022) collection.
- 95 2) Summer season (5/2022, 6/2022 and 7/2022) collection.
- 3) Autumn season (8/2022, 9/2022 and 10/2022) collection.
- 4) Winter (11/2022, 12/2022 and 1/2023) collection.

100 **Bacterial identification and counting:** Semen samples were cultured for bacterial detection and then incubated at 37 °C in aerobic and anaerobic conditions for 24-48 hours. Bacterial isolation and identification were conducted following Quinn *et al.* (2006). SAS (2012) analysis system was conducted to perform Chi-square test to detect the variation between the groups.

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Results

110 This study indicated that semen samples showed contamination with bacteria during winter and summer seasons. In sheep, six positive isolations recorded, which represent (16.67%) of 36

samples, while the spring and autumn seasons showed five positive samples (13.89%) from total 36 semen samples (Table 1). No significant variances ($P>0.01$) for samples of sheep semen between among seasons (Table 1). Table (2) shows that goat semen samples had six positives during winter and summer which represent (16.67%) of total samples, while in spring appear to result five positive samples (13.89%). Finally, autumn showed 3 positive samples (8.33%) of 36 tested samples. There was no significant differences ($P>0.05$) between all seasons (Table 2). Total percentage of positive samples for all the seasons in sheep and goat show that the sheep had more contaminated semen 22 of 144 (15.28%) samples as comparative with goat 20 of 144 (13.89%) (Table 1 and 2). There were no significant differences ($P>0.05$) between total semen contaminations between sheep and goat (Table 1 and 2). In contaminated sheep semen, isolated bacteria types identified are: *Klebsiella pneumonia*, *Proteus mirabilis*, *Escherichia coli* and *Staphylococcus aureus* (Table 3), whereas in goat semen goat are: *Staphylococcus aureus*, *Streptococcus faecalis*, *Pseudomonas aeruginosa*, *E. coli* and *Proteus mirabilis* (Table 4). The single isolation samples from sheep were 9 with 13 mixed contaminated samples, as well as 12 single and 8 mixed contaminated goat semen (Table 3 and 4).

Table 1: the number of bacterial isolation samples, positive isolation and percentage during different seasons of sheep ram semen samples.

Seasons	Samples number	Positive isolation	Percentage %	P value
Winter	36	6	16.67%	0.973*
Spring	36	5	13.89%	
Summer	36	6	16.67%	
Autumn	36	5	13.89%	
Total	144	22	15.28%	

* insignificant (P>0.05).

Table 2: the number of bacterial isolation samples, positive isolation and percentage during different seasons of goat buck semen samples.

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Seasons	Samples number	Positive isolation	Percentage %	P value
Winter	36	6	16.67%	0.503*
Spring	36	5	13.89%	
Summer	36	6	16.67%	
Autumn	36	3	8.33%	
Total	144	20	13.89%	

* insignificant (P>0.05).

Table 3: types of isolated bacteria from semen during different seasons in sheep ram.

Seasons	Bacterial isolated types	Single or mixed isolation
Winter	<i>Klebsiella pneumonia</i>	3 single
	<i>Proteus mirabilis</i>	3 mixed
Spring	<i>Klebsiella pneumonia</i>	2 single
	<i>Proteus mirabilis</i>	3 mixed
Summer	<i>Escherichia coli</i>	2 single
	<i>Staphylococcus aureus</i>	4 mixed
Autumn	<i>Escherichia coli</i>	2 single
	<i>Staphylococcus aureus</i>	3 mixed

Total	<i>Klebsiella pneumonia</i>	9 single 13 mixed
	<i>Proteus mirabilis</i>	
	<i>Escherichia coli</i>	
	<i>Staphylococcus aureus</i>	

Table 4: types of isolated bacteria from semen during different seasons in goat buck.

Seasons	Bacterial isolated types	Single or mixed isolation
Winter	<i>Escherichia coli</i>	4 single
	<i>Pseudomonas aeruginosa</i>	2 mixed
Spring	<i>Escherichia coli</i>	2 single
	<i>Staphylococcus aureus</i>	3 mixed
Summer	<i>Streptococcus faecalis</i>	3 single
	<i>Staphylococcus aureus</i>	3 mixed
Autumn	<i>Proteus mirabilis</i>	3 single
Total	<i>Escherichia coli</i>	12 single 8 mixed
	<i>Pseudomonas aeruginosa</i>	
	<i>Streptococcus faecalis</i>	
	<i>Staphylococcus aureus</i>	
	<i>Proteus mirabilis</i>	

Discussion

140 This study is the first trial to investigate the semen contamination in four consecutive seasons. Results indicated that sheep semen are contaminated with 15.28% while in goat's its 13.89%, but sheep's semen are contaminated with four different bacterial types compared to five different bacterial isolates in goat semen. There was no perfect ability to determine the way of the bacterial contamination in an ejaculate (Azawi and Ismaeel, 2011). Bacteria types that are mostly
145 isolated were *Staphylococcus aureus*, *Enterobacter cloacae*, *Proteus mirabilis*, *Staphylococcus epidermis*, and *Escherichia coli*, species. These five bacterial types represent 97% of all contaminated samples as mentioned by Yaniz *et al.* (2010), which agree with this study in regard to sheep and partially in regard to goat's contaminated semen. During storage, samples with less than 100 colony-forming units (CFU)/ml enterobacteria count had faster progressive sperm
150 motility with higher velocity at different times (Yaniz *et al.*, 2010). *E. coli* contaminated semen during storage showed downgrade in their viability, motility and velocity (Yaniz *et al.*, 2010). Sheep's semen with enterobacterial species contamination seemed lower in spermatozoa quality during cooling at 15°C (Yaniz *et al.*, 2010). This risk becomes true as the samples of our study became contaminated with such bacterial species. There was a statistical decrease ($P < 0.01$) in the
155 mean of number of live bacteria during middle of winter extended into spring ending, January showed lower of bacterial count (60.5 ± 2.98) with season significant effect in Awassi ram semen on bacterial number (Azawi and Ismaeel, 2011). This agrees partially with this study results. Bacterial presence role within semen has a wide disagreement opinion, so it is difficult to determine the cause of such contamination. Bacterial presence with semen, surly indicates
160 infection, due to transmission of this infection to the genital organs which can occur (Sanocka-Maciejewska *et al.*, 2005). Sperm morphology was affected with inflammatory cells in some cases of bacterial contamination, and bacterial presence in semen ejaculates affected directly on

fertilization of ova (Fraczek and Kurpisz, 2015), by attaching with spermatozoa, decreasing their motility and inducing acrosome reaction (Azawi and Ismaeel, 2011). Microbes affect indirectly through toxins production (Wang *et al.*, 2021). Semen is considered as a good quality when there is low bacterial contains (Azawi and Ismaeel, 2011). Semen viability decreases within a short time if their sperms are contaminated, resulting sperms death leading to the risk of increasing female reproductive tract pathologies, and eventually, decreasing fertility (Azawi and Ismaeel, 2011). This fact shows the importance of detecting any bacterial contamination of semen before using it in breeding. Some bacterial types presented in the genital system of the females without any effect upon the function of reproduction (Zaid, 2009). Zaid (2009) determine the bacterial microflora isolation inside ovine vagina between 56.5-96.5%, while in the caprine (71.4%) has been reported by Al-Delemi (2005) during diestrous phase. Another factor can be the male external genital parts contamination during mating with female. The isolation of more than one bacterium in one swabs was identify by Zaid (2009). This fact is similar to our results. In the ewe percentage of large bacteria count which had been isolated as *Escherichia coli* and Enterobacter spices with some other bacteria. This agrees with other researchers results found before (Al-Delemi, 2005). Otherwise, the does had highest percentage isolates with *Streptococcus faecalis* and *Pseudomonas aeruginosa* (Zaid, 2009) which agrees with this study. This study revealed that the external parts of genital system of rams were carrying many bacterial species. This finding agrees with other authors (Zaid and Al-Zubaidy, 2009 and Pohjanvirta *et al.*, 2020). The resent results revealed that the bacteria which cultured from male semen were partly similar to other authors (Zaid *et al.*, 2007). A normal microflora contains many bacterial types can grow rapidly and decrease body immunity causing diseases, while there are other which are considered as pathogenic types (Zaid and Al-Zubaidy, 2009). Viability of sperm fertilization with bacterial number and bacterial types is an important relationship (Zaid and Al-Zubaidy, 2009). However Zaid and Al-Zubaidy (2009) found that there were ten folds little bacterial count in the second

ejaculation compared to the first. This result may be explaining the decrease of contamination percentage during breeding season of goats in our research. The recent study showed that it can be culture more than one type of bacteria from the same swab; this fits with other authors (Al-Delemi, 2005). Normal male semen contains *Staphylococcus aureus* with large amount (Zaid and Al-Zubaidy, 2009). This agrees with our study. After mating *Pseudomonus aeruginosa* isolated by (Al-Delemi, 2005) and it is considered as normal flora in ewes. In this study the isolation of such bacterium is done in male goats. Semen bacterium refers to low fertility; also, there was a relationship with male sterility (Zaid and Al-Zubaidy, 2009). The contamination of semen with *Esherichia coli* or *Pseudomonus aeruginosa* may be resulted from feces presence during mating of the male (Zaid and Al-Zubaidy, 2009). There was a seasonal effect on semen quality in different animal species (Ibrahim and Zaid, 2015; Zaid, 2017; Ibahim and Zaid, 2017a; 2017b). In this part these studies did not deal with bacterial contamination as that which done by our study.

Conclusion

the semen of sheep and goat had contaminated with normal flora bacteria out of breeding uses, while the seasons had no effect on this bacterial contamination.

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References

- Al-Delemi, DHJ. (2005) The normal bacterial flora in the vaginal cavity of Iraqi cows, sheeps, goats and camels during the luteal phase. **Al-Qadisiya Journal of Veterinary Medicine Science**, 4 (1): 23-29.
215 https://www.researchgate.net/publication/308780835_The_Normal_Bacterial_Flora_in_the_Vaginal_Cavity_of_Iraqi_Cows_Sheep's_Goats_and_Camels_During_the_Luteal_Phase
- Al-Tae, HSR., Al-Samarrae, IAA., Al-Ahmed, HI. (2019) Antibiotic Susceptibility and Molecular Detection of Pseudomonas aeruginosa Isolated from Bovine Mastitis. **Iraqi Journal of Veterinary Medicine**, 43(2): 81-89.
220 <https://doi.org/10.30539/iraqijvm.v43i2.536>
- Al-Taii, DHF., Yousif, AA. (2019) Effects of E.coli O157:H7 Experimental Infections on Rabbits. **Iraqi Journal of Veterinary Medicine**, 43(1): 34-42.
<https://doi.org/10.30539/iraqijvm.v43i1.468>
- Anvar, SAA., Nowruai, B., Afshari, G. (2023). A Review of the Application of Nanoparticles Biosynthesized by Microalgae and Cyanobacteria in Medical and Veterinary Sciences. **Iranian Journal of Veterinary Medicine**, 17 (1): 1-18. [DOI: 10.22059/IJVM.17.1.1005309](https://doi.org/10.22059/IJVM.17.1.1005309)
225
- Azawi, OL., Ismaeel, MA. (2011) Effects of Seasons on Some Semen Parameters and Bacterial Contamination of Awassi ram Semen. **Reproduction in Domestic Animal**, 47 (3): 403-406. [doi: 10.1111/j.1439-0531.2011.01888.x](https://doi.org/10.1111/j.1439-0531.2011.01888.x)
- 230 Enwuru, CA., Iwalokun, B., Enwuru, VN., Ezechi, O., Oluwadun, A. (2016) The effect of presence of facultative bacteria species on semen and sperm quality of men seeking

fertility care. **African Journal of Urology**, 22(3): 213-222.
<https://doi.org/10.1016/j.afju.2016.03.010>

235 **Foroutan, S., Eslampour, MA., Emaneini, M., Jabalameli, F., Akbari, G. (2022).**
Characterization of Biofilm Formation Ability, Virulence Factors and Antibiotic
Resistance Pattern of Staphylococcus aureus Isolates from Subclinical Bovine Mastitis.
Iranian Journal of Veterinary Medicine, 16 (2): 144-154. [DOI:
10.22059/IJVM.2021.323994.1005174](https://doi.org/10.22059/IJVM.2021.323994.1005174)

240 **Fraczek, M., Kurpysz, M., (2015)** Mechanisms of the harmful effects of bacterial semen
infection on ejaculated human spermatozoa: potential inflammatory markers in semen.
Folia Histochem Cytobiol, 53(3): 201-217. [doi: 10.5603/fhc.a2015.0019](https://doi.org/10.5603/fhc.a2015.0019)

245 **Gaddafi, MS., yakubu, Y., Junaidu, AUU., Bello, MB., Bitrus, AA., Musawa, AI., Garba,
B., Lawal, H. (2023).** Occurrence of Methicillin-resistant Staphylococcus aureus (MRSA)
From Dairy Cows in Kebbi, Nigeria. **Iranian Journal of Veterinary Medicine**, 17 (1):
19-26. [DOI: 10.22059/IJVM.17.1.1005256](https://doi.org/10.22059/IJVM.17.1.1005256)

Gharban, HAJ and Yousif, AA (2020). Serological and Molecular Phylogenetic Detection of
Coxiella burnetii in Lactating Cows, Iraq. **Iraqi Journal of Veterinary Medicine**,
44(E0): 42-50. [https://doi.org/10.30539/ijvm.v44i\(E0\).1020](https://doi.org/10.30539/ijvm.v44i(E0).1020)

250 **Hanoun, AT., Al-Samrrae, IAA. (2019)** Isolation and Identification of Escherichia coli and
Salmonella typhimurium from Sheep in Baghdad city. **Iraqi Journal of Veterinary
Medicine**, 43(1): 124-129. <https://doi.org/10.30539/iraqijvm.v43i1.482>

Hobson, N., Chousalkar, KK., Chenoweth, PJ. (2013) Ureaplasma diversumin bull semen in
Australia: its detection and potential effects. **Australian Veterinary Journal**, 91(11): 469-
473. [doi: 10.1111/avj.12113](https://doi.org/10.1111/avj.12113)

- 255 **Ibrahim, NS., Zaid, NW. (2015).** Effect of the Season on Male Dog Testosterone, SSH, LH Level in Iraq. **Indian Journal of Research**, 4 (9): 10-11. DOI: https://www.worldwidejournals.com/paripex/recent_issues_pdf/2015/August/August_2015_1439551076__112.pdf.
- Ibrahim, NS., Zaid, NW. (2017a).** Testosterone role during seasons changes in the dogs testes. **The Iraqi Journal of Veterinary Medicine**, 41 (1): 125–130. DOI: <https://doi.org/10.30539/iraqijvm.v41i1.93>
- 260 **Ibrahim, NS., Zaid, NW. (2017b).** Semen evaluation in local dogs during different season in Baghdad. **Iraqi Journal of Veterinary Science**, 31 (1): 45-50. DOI: [10.33899/IJVS.2017.126719](https://doi.org/10.33899/IJVS.2017.126719)
- Karashahin, T., Dursun, S., Aksoy, NH., Ipek, H., Senturk, G. (2023).** Hematological Parameters in Hair Goats During and out of Breeding Season Hair Goats Seasonal Hematological Parameters. **Iranian Journal of Veterinary Medicine**, 17 (2): 113-118. DOI: [10.32598/IJVM.17.2.1005334](https://doi.org/10.32598/IJVM.17.2.1005334)
- 265 **Luther, A., Beckermann, C., Nguyen, TQ., Verspohl, J., Waberski, D. (2023)** Growth Dynamic and Threshold Values for Spermicidal Effects of Multidrug-Resistant Bacteria in Extended Boar Semen. **Microorganisms** 11(3): 788. <https://doi.org/10.3390/microorganisms11030788>
- 270 **Meena, GS., Bhakat, M., Raina, VS., Gupta, AK., Mohanty, TK., Bishist, R. (2017)** Effect of different antibiotic combinations in extender on bacterial load and seminal characteristics of Murrah bulls. **Buffalo Bulletin**, 36(1): 251-257. DOI: <https://kuojs.lib.ku.ac.th/index.php/BufBu/article/view/1240/332>
- 275 **Meena, GS., Raina, VS., Gupta, AK., Mohanty, TK., Bhakat, M., Abdullah, M., Bishist, R. (2015)** Effect of preputial washing on bacterial load and preservability of semen in Murrah buffalo bulls. **Veterinary World**, 8(6): 798–803. doi: [10.14202/vetworld.2015.798-803](https://doi.org/10.14202/vetworld.2015.798-803)
- 280 **Mitra, J., Chowdhury, S., Panda, S., Chakraborty, M., Singha, A. (2016)** Microbiological evaluation of bovine frozen semen samples in west bengal, India, 2016. **Exploratory**

Animal and Medical Research, 6(2): 185-191.

https://www.animalmedicalresearch.org/Vol.6_Issue-2_December_2016/Joyjirt%20Mitra.pdf

Morrell, JM., Wallgren, M. (2014) Alternatives to Antibiotics in Semen Extenders: A Review. **Pathogens, 3: 934-946.** [doi: 10.3390/pathogens3040934](https://doi.org/10.3390/pathogens3040934)

285 **Noakes, DE., Parkinson, TJ., England, GCW. (2019)** Veterinary Reproduction and Obstetrics. 10th Ed. Elsevier Sci. p. 408-419. <https://www.elsevier.com/books/veterinary-reproduction-and-obstetrics/noakes/978-0-7020-7233-8>

Pohjanvirta, T., Vahanikkila, N., Simonen, H., Pelkonen, S., Autio, T. (2020) Efficacy of Two Antibiotic-Extender Combinations on Mycoplasma bovis in Bovine Semen
290 Production. **Pathogens, 9(10): 808.** <https://doi.org/10.3390/pathogens9100808>

Quinn, PJ; Markey, BK; Carter, ME; Donnelly, WJC; Leonard, FC and Maghire, D (2006). Veterinary microbiology and microbial diseases. 6th ed. Black well Publishing Company, Great Britain. <https://www.wiley.com/en-be/Veterinary+Microbiology+and+Microbial+Disease%2C+2nd+Edition-p-9781405158237>

295 **Razook, BRF; Al-Ani, AN and Mahmood, MM (2020).** Hematological Picture of Rabbits Immunized with Pseudomonas aeruginosa. **Iraqi Journal of Veterinary Medicine, 44(E0): 64-68.** [https://doi.org/10.30539/ijvm.v44i\(E0\).1023](https://doi.org/10.30539/ijvm.v44i(E0).1023)

Reda, AA., Almaw, G., Abreha, S., Tadeg, W., Tadesse, B. (2020) Bacteriospermia and Sperm Quality of Cryopreserved Bull Semen Used in Artificial Insemination of Cows in
300 South Wollo Zone, Ethiopia. **Veterinary Medicine International, 2020: 2098315.** [doi: 10.1155/2020/2098315](https://doi.org/10.1155/2020/2098315)

Russell, PH., Lyaruu, VH., Millar, JD., Curry, MR., Watson, PF. (1997) The potential transmission of infectious agents by semen packaging during storage for artificial insemination. **Animal Reproduction Science, 47(4): 337-342.** [doi: 10.1016/s0378-4320\(97\)00017-1.](https://doi.org/10.1016/s0378-4320(97)00017-1)
305

- Sannat, C., Nair, A., Sahu, SB., Sahasrabudhe, SA., Kumar, A., Gupta, AK., Shende, RK.** (2015) Effect of species, breed, and age on bacterial load in bovine and bubaline semen. **Veterinary World**, 8(4): 461-466. [doi: 10.14202/vetworld.2015.461-466](https://doi.org/10.14202/vetworld.2015.461-466)
- 310 **Sanocka-Maciejevska, D; Ciupinska, M and Kurpisz, M (2005).** Bacterial infection and semen quality. **Journal of Reproductive and Immunology**, 67 (1-2): 51-56. <https://doi.org/10.1016/j.jri.2005.06.003>
- SAS (2012).** Statistical Analysis System, Usre's Guide. Statistical Version 9.1th ed. SAS. Inst. Inc. Cary. N.C. USA.
- 315 **Schulze, M., Nitsche-Melkus, E., Hensel, B., Jung, M., Jakop, U. (2020)** Antibiotics and their alternatives in Artificial Breeding in livestock. **Animal Reproduction Science**, 2020: 106284 <https://doi.org/10.1016/j.anireprosci.2020.106284>
- Thacker, BJ., Larsen, RE., Joo, HS., Leman, AD. (1984)** Swine diseases transmissible with artificial insemination. **Journal of the American Veterinary Medical Association**, 185(5): 511-517. [PMID: 6090372](https://pubmed.ncbi.nlm.nih.gov/6090372/)
- 320 **Thibier, M., Guerin, B. (2000)** Hygienic aspects of storage and use of semen for artificial insemination. **Animal Reproduction Science**, 62: 233-251. [DOI:10.1016/S0378-4320\(00\)00161-5](https://doi.org/10.1016/S0378-4320(00)00161-5)
- 325 **Wang, S., Zhang, K. Yao, Y. Li, J., Deng, S. (2021)** Bacterial Infections Affect Male Fertility: A Focus on the Oxidative Stress-Autophagy Axis. **Frontiers in Cell and Developmental Biology**, 9: <https://doi.org/10.3389/fcell.2021.727812>
- Yaniz; JL., Marco-Aguado; MA., Mateos; JA., Santolaria; P. (2010)** Bacterial contamination of ram semen, antibiotic sensitivities, and effects on sperm quality during storage at 15°C. **Animal Reproduction Science**, 122 (1-2): 142-149. [doi: 10.1016/j.anireprosci.2010.08.006](https://doi.org/10.1016/j.anireprosci.2010.08.006)

- 330 **Zaid, NW. (2009)** Vaginal flora of Iraqi sheep and goats during different reproductive stages.
Al-Anbar Journal of Veterinary Sciences, 2 (1): 25-30. <https://www.iasj.net/iasj/article/30719>
- Zaid, NW. (2017).** Seasonal changes on epididymal histology and testosterone receptors in Iraqi dogs. **Bulletin of the Iraqi Natural History Museum**, 14 (4): 275-284. DOI: <https://doi.org/10.26842/binhm.7.2017.14.4.0275>
- 335 **Zaid, NW., Abdalla, AH., Ibrahim, AH., Khadim, AA. (2007)** Effect of prostoglandine PGF2 α on bacterial elimination during puerperium in Awassi ewes. **Al-Anbar Journal of Agriculture Science**, 5 (1): 211-217. DOI: [10.32649/ajas.2007.36264](https://doi.org/10.32649/ajas.2007.36264)
- Zaid, NW., Al-Zubaidy, IA. (2009)** The Effect of Natural Mating on the Bacterial Pollution in the Endogenous Ram. **Al-Anbar Journal of Veterinary Sciences**, 2 (1): 31-35. <https://www.iasj.net/iasj/article/30720>
- 340